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# (54) POLYPEPTIDES HAVING TRANSGALACTOSYLATING ACTIVITY

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	A23C 9/00	(2006.01)
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	A23C 19/032	(2006.01)
	A23C 19/05	(2006.01)
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(52) U.S. Cl.

#### (58) Field of Classification Search

CPC	A23C 9/00
USPC	435/42, 43, 522
See application file for complete sear	ch history.

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# (57) ABSTRACT

The present invention relates to polypeptides, specifically polypeptides having transgalactosylating activity and nucleic acids encoding these, and their uses in e.g. dairy product.

# 3 Claims, 4 Drawing Sheets

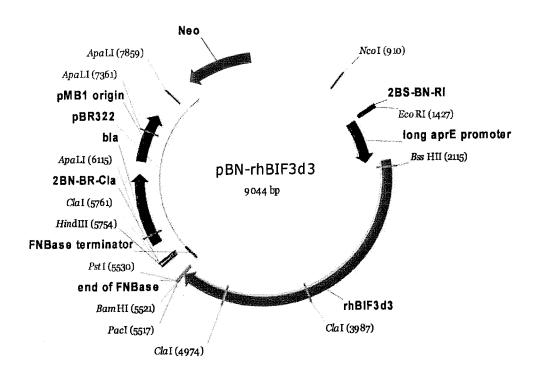
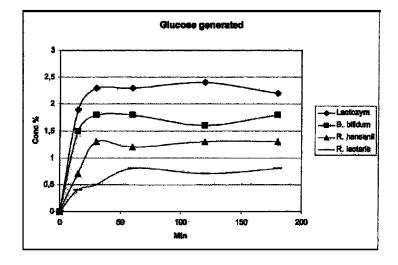


Fig. 1



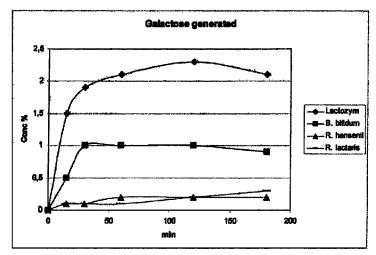
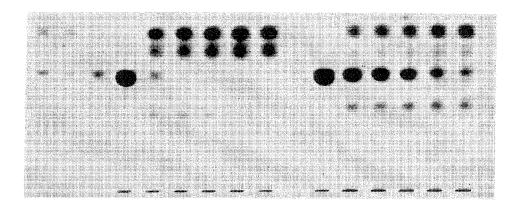


Fig. 2

10 11 12 13 14 15



16 17 18 19 20 21 26 27

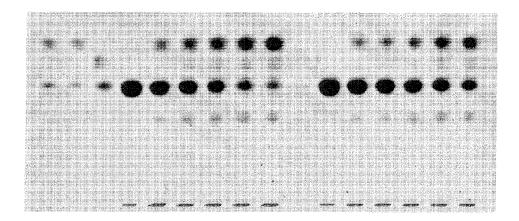


Fig. 3

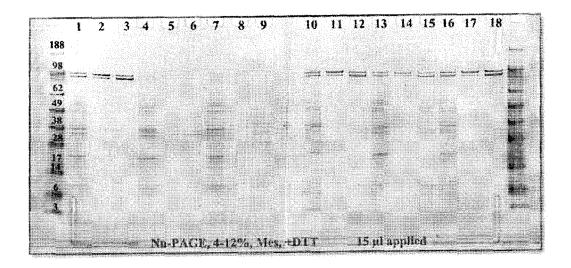


Fig. 4

# POLYPEPTIDES HAVING TRANSGALACTOSYLATING ACTIVITY

## CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims priority to PCT Application No. PCT/IB2011/054865 entitled "Polypeptides Having Transgalactosylating Activity," filed Mar. 29, 2011, which claims priority to U.S. Provisional Application No. 61/318,580, filed 10 Mar. 29, 2010 and EP No. 10158172.6, filed Mar. 29, 2010, all of which are expressly incorporated by reference herein in their entirety.

#### SEQUENCE LISTING

A text file in compliance with ASCII and having a ".txt" extension has been electronically submitted via EFS-Web. The text file named "Sequence Listing" was created on Nov. porated by reference herein in its entirety.

# FIELD OF THE INVENTION

The present invention relates to polypeptides, specifically 25 polypeptides having transgalactosylating activity and nucleic acids encoding these, and their uses in e.g. dairy product.

#### BACKGROUND OF THE INVENTION

Galactooligosaccharides (GOS) are carbohydrates which are nondigestable in humans and animals comprising two or more galactose molecules, typically up to nine, linked by glycosidic bonds. GOS's may also include one or more glucose molecules. One of the beneficial effects of GOS's is their 35 ability of acting as prebiotic compounds by selectively stimulating the proliferation of beneficial colonic microorganisms such as bacteria to give physlologicai benefits to the consumer. The established health effects have resulted in a growing interest In GOSs as food ingredients for various types of 40 food.

The enzyme  $\beta$ -galactosidase (EC 3.2.1.23) usually hydrolyses lactose to the monosaccharides D-glucose and D-galactose. In the normal enzyme reaction of β-galactosidases, the enzyme hydrolyses lactose and transiently binds 45 the galactose monosaccharide in a galactose-enzyme complex that transfers galactose to the hydroxyl group of water, resulting in the liberation of D-galactose and D-glucose. However, at high lactose concentrations some  $\beta$ -galactosidases are able to transfer galactose to the hydroxyl groups of 50 D-galactose or D-glucose in a process called transgalactosylation whereby galacto-oligosaccharides are produced.

The genus *Bifidobacterium* is one of the most commonly used types of bacteria cultures in the dairy industry for fermenting a variety of diary products. Ingestion of Bifidobac- 55 terium-containing products furthermore has a health-promoting effect. This effect is not only achieved by a lowered pH of the intestinal contents but also by the ability of Bifidobacterium to repopulate the intestinal flora in individuals who have had their intestinal flora disturbed by for example intake of 60 antibiotics. Bifidobacterium furthermore has the potential of outcompeting potential harmful intestinal micro-organisms.

Galacto-oligosaccharides are known to enhance the growth of Bifidobacterium. This effect is likely achieved through the unique ability of Bifidobacterium to exploit 65 galacto-oilgosaccharides as a carbon source. Dietary supplement of dalacto-oligosaccharides is furthermore thought to

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have a number of long-term disease protecting effects. For example, galacto-oligosaccharide intake has been shown to be highly protective against development of colorectal cancer in rats. There is therefore a great interest in developing cheap and efficient methods for producing galacto-oligosaccharides for use in the industry for improving dietary supplements and dairy products.

A beta-galactosidase polypeptide from Ruminococcus/ Blautia hansenii having 1807 amino acids (having SEQ ID NO: 12) is known from the database UniProt, 24 Nov. 2009, Full=Beta-galactosidase" XP002591904 retrieved from EBI accession no. UNIPROT:C9LAL1.

A glycosidase having 1768 amino acids (having SEQ ID NO: 13) is known from the database UniProt, 14 Oct. 2008, "Subname: uncharacterised Full=Putative XP002610554 retrieved from EBI accession no. UNIPROT: B5CQV4.

An extracellular lactase from Bifidobacterium bificium 7, 2012 and is 87,924 bytes. The text file is expressly incor- 20 DSM20215 truncated with approximately 580 amino acids (BIF3) has been described as a transgalactosylating enzyme in a solution containing lactose solubilised in water (Jørgensen et al. (2001), Appl. Microbiol. Biotechnol., 57: 647-652). In WO 2009/071539 a differently truncated fragment compared to BIF3 is described as resulting in efficient hydrolysis and very low production of GOS when tested in

> The Bifidobacterium bifidum lactase enzymes described above have the drawback of either requiring high lactose concentrations in order to exhibit transgalactosylase activity or predominantly having beta-galactosylase (hydrolase) activity.

> There is still a need to develop enzymes that are efficient at producing GOS and which furthermore can work at low lactose substrate levels such as in milk.

# OBJECT OF THE INVENTION

It is an object of embodiments of the invention to provide a polypeptide which has a useful ratio of transgalactosylation to hydrolysis activity and thus are efficient producers of GOS when incubated with lactose even at low lactose levels such as in a milk-based product. It is a further object of embodiments of the invention to provide a method for production of galacto-oligosaccharides (GOS) in situ in dairy products. It is a further object of embodiments of the invention to provide a method for developing a cheaper and more efficient method for production of galacto-oligosaccharides (GOS) for use in the industry.

# SUMMARY OF THE INVENTION

The present invention discloses two related polypeptides, which surprisingly are able to produce galacto-oligosaccharidesin situ when incubated with lactose such as milk. Thus, when the polypeptide, as described herein, or a host cell expressing the polypeptide is incubated with lactose under appropriate conditions, galacto-oligosaccharides are produced at a high efficiency and thus lactose is reduced. The presence of galacto-oligosaccharides diary products or other comestible products has the advantage of enhancing the growth of healthpromoting Bifdobacterium sp. in the product or in the intestinal flora of the consumer after intake of the product or both.

In one aspect, the invention relates to an isolated polypeptide having transgalactosylating activity selected from the group consisting of:

- a. a polypeptide comprising an amino acid sequence having at least 66% sequence identity to the amino acid sequence of the mature polypeptide of SEQ ID NO: 1,
- b. a polypeptide comprising an amino acid sequence having at least 66% sequence identity to the amino acid sequence of the mature polypeptide of SEQ ID NO: 2,
- c. a polypeptide encoded by a polynucleotide that hybridizes under at least low stringency conditions with i) the nucleic acid sequence comprised in SEQ ID NO: 10 encoding the mature polypeptide of SEQ ID NO: 1; ii) 10 the cDNA sequence of i) or iii) the complementary strand of i) or ii);
- d. a polypeptide encoded by a polynucleotide that hybridizes under at least low stringency conditions with i) the nucleic acid sequence comprised in SEQ ID NO: 11 15 having encoding the mature polypeptide of SEQ ID NO: 2; ii) a. at the cDNA sequence of i) or iii) the complementary strand of i) or iii;
- e. a polypeptide comprising a conservative substitution, deletion and/or insertion of one or more amino acids of 20 SEQ ID NO: 1, and
- f. a polypeptide comprising a conservative substitution, deletion and/or insertion of one or more amino acids of SEO ID NO: 2.

provided that the polypeptide of above items a, c, and e at the 25 most has a length of 1806 amino acids and provided that the polypeptide of above items b, d, and f at the most has a length of 1767 amino acids.

In one aspect, disclosed herein is a method of expressing a polypeptide, the method comprising obtaining a cell as dis-30 closed herein and expressing the polypeptide from the cell, and optionally purifying the polypeptide. In a further aspect, disclosed herein is a composition comprising a polypeptide as disclosed herein, preferably a food composition, more preferably a dairy product. In a further aspect, disclosed herein is 35 a method for producing a food product by treating a substrate comprising lactose with a polypeptide as disclosed herein such as producing a dairy product by treating a milk-based substrate comprising lactose with a polypeptide as disclosed herein. In a further aspect, the polypeptides are used for 40 treating a substrate with a hydrolysing beta-galactosidase. In a further aspect, disclosed herein is a food product, preferably a dairy product, comprising a transgalactosylating enzyme obtained from Ruminococcus hansenii or Ruminococcus lactaris, preferably as defined in item a-f in above, and more 45 preferably a polypeptide as further defined herein. In yet an aspect, disclosed herein is a galacto-oligosaccharide or composition thereof obtained by treating a substrate comprising lactose with a polypeptide as disclosed herein.

In one aspect, a polypeptide having transoalactosylating 50 activity comprising an amino add sequence having

- a. at least 66% sequence identity to the amino acid sequence of SEQ ID NO: 1, and/or
- b. at least 66% sequence identity to the amino acid sequence of SEQ ID NO: 2, is provided.

In another aspect, a polypeptide having a ratio of transgalactosylting activity:  $\beta$ -galactosidase activity of at least 1 as measured at a concentration of 6 LAU/ml in a milk-based assay at 37° C. and 5 w/w % lactose after 30 minutes reaction comprising an amino add sequence having

- a. at least 66% sequence identity to the amino acid sequence of SEQ ID NO: 1, and/or
- b. at least 66% sequence identity to the amino acid sequence of SEQ ID NO: 2, is provided.

In a further aspect, a polypeptide comprising an amino acid 65 sequence having at least 60% sequence identity to the amino acid sequence of SEQ ID NO: 5, is provided. In a further

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aspect, a polypeptide comprising an amino acid sequence having at least 94% sequence identity to the amino acid sequence of SEQ ID NO: 8, is provided. In a further aspect, the use of a polypeptide having transgalactosylating activity comprising an amino acid sequence having at least 60% sequence identity to the amino acid sequence of SEQ ID NO: 5 for producing galacto-oligosaccharides, is provided. In a further aspect, the use of a polypeptide having transgalactosylating activity comprising an amino acid sequence having at least 94% sequence identity to the amino acid sequence of SEQ ID NO: 8 for producing galacto-oligosaccharides, is provided.

In a further aspect, the use of a polypeptide having transgalactosylating activity comprising an amino acid sequence having

- a. at least 60% sequence identity to the amino acid sequence of SEQ ID NO: 3,
- b. at least 60% sequence identity to the amino acid sequence of SEQ ID NO: 4,
- c. at least 60% sequence identity to the amino acid sequence of SEQ ID NO: 6, or
- d. at least 80% sequence identity to the amino acid sequence of SEQ ID NO: 7, for producing galacto-oligosaccharides, is provided.

In a further aspect, a polypeptide having a ratio of transgalactosylating activity:β-galactosidase activity of at least 1 as measured at a concentration of 6 LAU/ml in a milk-based assay at 37° C. and 5 w/w % lactose after 30 minutes reaction, is provided. In a further aspect, a nucleic acid capable of encoding a polypeptide as disclosed herein, is provided. In a further aspect, a plasmid comprising a nucleic acid as disclosed herein, is provided. In a further aspect, an expression vector comprising a nucleic acid as disclosed herein, or capable of expressing a polypeptide as disclosed herein, is provided. In a further aspect, a host cell comprising, preferably transformed with, a plasmid as disclosed herein, or an expression vector as disclosed herein, is provided. In a further aspect, a cell capable of expressing a polypeptide as disclosed herein, is provided. In a further aspect, a method of expressing a polypeptide, the method comprising obtaining a host cell or a cell as disclosed herein and expressing the polypeptide from the cell or host cell, and optionally purifying the polypeptide, is provided. In a further aspect, a composition comprising a polypeptide as disclosed herein and a stabilizer, is provided. In a further aspect, a composition comprising a polypeptide as disclosed herein and a carbohydrate substrate, is provided. In a further aspect, a method for producing a dairy product by treating a milk-based substrate comprising lactose with a polypeptide having a ratio of transgalactosylating activity:βgalactosidase activity of at least 1, at least 2.5, at least 3, at least 4, at least 5, at least 6, at least 7, at least 8, at least 9, at least 10, at least 11, or at least 12 as measured at a concentration of 6 LAU/ml in a milk-based assay at 37° C. and 5 w/w % lactose after 30 minutes reaction is provided. In a further 55 aspect, a method for producing a dairy product by treating a milk-based substrate comprising lactose with a polypeptide as disclosed herein, is provided. In a further aspect, a use of a cell as disclosed herein for producing a product selected from the group consisting of yoghurt, cheese, fermented milk prod-60 uct, dietary supplement and probiotic comestible product, is provided. In a further aspect, a dairy product comprising a cell as disclosed herein, is provided. In a further aspect, a dairy product comprising a polypeptide as disclosed herein, is provided. In a further aspect, a dairy product comprising a polypeptide as disclosed herein in a concentration of 001-1000 ppm, is provided. In a further aspect, a dairy product comprising an inactivated polypeptide as disclosed herein, is

provided. In a further aspect, a dairy product comprising GOS formed in situ by a polypeptide as disclosed herein, is provided. In a further aspect, a use of a transgalactosylating polypeptide as disclosed herein or a cell as disclosed herein for producing galacto-oligosaccharides, is provided. In a further aspect, a use of a transgalactosylating polypeptide as disclosed herein or a cell as disclosed herein, for producing galacto-oligosaccharides to be part of a product selected from the group consisting of yoghurt, cheese, fermented dairy products, dietary supplements and probiotic comestible products, is provided. In a further aspect, a use of a transgalactosylating polypeptide as disclosed herein or a cell as disclosed herein, for producing galacto-oligosaccharides to enhance the growth of Bifidobacterium, is provided. In a further 15 aspect, a use of a transgalactosylating polypeptide as disclosed herein or a cell as disclosed herein, for producing galacto-oligosaccharides to enhance the growth of Bifidobacterium in a mixed culture fermentation, is provided. In a further aspect, a process for producing a transgalactosylating 20 polypeptide as disclosed herein, comprising culturing a cell as disclosed herein in a suitable culture medium under conditions permitting expression of said polypeptide, and recovering the resulting polypeptide from the culture, is provided. In a further aspect, a process for producing galacto-oligosac- 25 charides, comprising contacting of an polypeptide as disclosed herein or a cell as disclosed herein with a milk-based solution comprising lactose.

#### SEQUENCE LISTING

SEQ ID NO: 1 is a 1125 amino acid truncated fragment of SEQ ID NO: 12.

SEQ ID NO: 2 is 1150 amino acid truncated fragment of SEQ ID NO: 13.

 $\dot{\rm SEQ}$  ID NO: 3 is amino acid residues 559-649 of SEQ ID No: 1.

SEQ ID NO: 4 is amino acid residues 579-649 of SEQ ID No: 1.

SEQ ID NO: 5 is amino acid residues 579-636 of SEQ ID No: 1.

SEQ ID NO: 6 is amino acid residues 575-665 of SEQ ID No: 2.

SEQ ID NO: 7 is amino acid residues 594-665 of SEQ ID 45 No: 2.

SEQ ID NO: 8 is amino acid residues 594-652 of SEQ ID No: 2.

SEQ ID NO: 9 is a signal peptide from the pBN *Bacillus subtilis* expression vector.

SEQ ID NO: 10 is the nucleotide sequence encoding SEQ ID NO: 1 including sequence encoding the signal peptide.

SEQ ID NO: 11 is the nucleotide sequence encoding SEQ ID NO: 2 including sequence encoding the signal peptide.

SEQ ID NO: 12 is a beta-galactosidase from *Ruminococ- cus/Blautia hansenii* DSM 20583.

SEQ ID NO: 13 is a glycosidase from *Ruminococcus lactaris*, ATCC 29176.

SEQ ID NO: 14 is the nucleotide sequence encoding SEQ  $_{60}$  ID NO: 12 without the signal sequence.

SEQ ID NO: 15 is the nucleotide sequence encoding SEQ ID NO: 13 without the signal sequence.

SEQ ID NO: 16 is the nucleotide sequence encoding SEQ ID NO: 1.

SEQ ID NO: 17 is the nucleotide sequence encoding SEQ ID NO: 2.

# LEGENDS TO THE FIGURE

FIG. 1 shows a plasmid map of the *Ruminococcus hansenii* expression construct. The rhBIF3d3 coding sequence was fused inframe with the aprE signal sequence using BssHII and PacI as restriction sites.

FIG. 2 shows accumulation of galactose and glucose during enzymatic treatment of a 5% w/w lactose solution in T-buffer with Lactozym® as control, *Ruminococcus hansenii* (SEQ ID NO: 1), *Ruminococcus lactaris* (SEQ ID NO:2) and *Bifidobacterium bifidum* BIF3d3 (truncated) (as described by Jørgensen et al. (2001), Appl. Microbiol. Biotechnol., 57: 647-652 and EP patent 1,283,876).

FIG. 3 shows the result of Thin Layer Chromatography of the polypeptides in 9 w/w % reconstituted milk giving a final concentration of lactose of 5% w/w. The polypeptides were dosed based upon the LAU activity determined as described in example 1 at a final concentration of 6 LAU/ml.

FIG. 4 shows the results of the anion exchange chromatography of variants of the *Ruminococcus hansenii* (SEQ ID NO:1). The gel is a Nu-PAGE 4-12% acrylamide gel stained with coomassie brilliant blue staining.

#### DETAILED DISCLOSURE OF THE INVENTION

Disclosed herein is an isolated polypeptide having transgalactosylating activity selected from the group consisting of:

- a. a polypeptide comprising an amino acid sequence having at least 66% sequence identity to the amino acid sequence of the mature polypeptide of SEQ ID NO: 1,
- a polypeptide comprising an amino acid sequence having at least 66% sequence identity to the amino acid sequence of the mature polypeptide of SEQ ID NO: 2,
- c. a polypeptide encoded by a polynucleotide that hybridizes under at least low stringency conditions with i) the nucleic acid sequence comprised in SEQ ID NO: 10 encoding the mature polypeptide of SEQ ID NO: 1; ii) the cDNA sequence of i) or iii) the complementary strand of i) or ii);
- d. a polypeptide encoded by a polynucleotide that hybridizes under at least low stringency conditions with i) the nucleic acid sequence comprised in SEQ ID NO: 11 encoding the mature polypeptide of SEQ ID NO: 2; ii) the cDNA sequence of i) or iii) the complementary strand of i) or ii);
- e. a polypeptide comprising a conservative substitution, deletion and/or insertion of one or more amino acids of SEQ ID NO: 1, and
- f. a polypeptide comprising a conservative substitution, deletion and/or insertion of one or more amino acids of SEQ ID NO: 2,

provided that the polypeptide of above items a, c, and e at the most has a length of 1806 amino acids and provided that the polypeptide of above items b, d, and f at the most has a length of 1767 amino acids.

In accordance with this detailed description, the following abbreviations and definitions apply; It should be noted that as used herein, the singular forms "a," "an," and "the" include plural referents unless the context clearly dictates otherwise. Thus, for example, reference to "an polypeptide" includes a plurality of such polypeptides, and reference to "the formulation" includes reference to one or more formulations and equivalents thereof known to those skilled in the art, and so forth.

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art. The following terms are provided below.

"Transgalactosylase" means an enzyme that, among other 5 things, is able to transfer galactose to the hydroxyl groups of D-galactose or D-glucose whereby galacto-oligosaccharides are produced. In one aspect, a transgalactosylase is identified by reaction of the enzyme on lactose in which the amount of galactose generated is less than the amount of glucose generated at any given time.

In the present context, the term "transgalactosylating activity" means the transfer of a galactose moiety to a molecule other than water and is measured as [glucose]–[galactose] generated at any given time during reaction.

In the present context the term " $\beta$ -galactosidase activity" means the ability of an enzyme to hydrolyse  $\beta$ -galactosides such as for example lactose into monosaccharides, glucose and galactose.

In the present context, the term "relative transgalactosylation activity" means ([Glucose]-[Galactose]100)/[Glucose]) measured at a timepoint after 15 minutes of reaction time.

In the present context, the term [Glucose] means the glucose concentration in % by weight as measured by HPLC.

In the present context, the term [Galactose] means the 25 galactose concentration in % by weight as measured by HPLC.

In the present context, the term "after 15 min. reaction" means the amount of time which should pass before measurement of activity after incubation with the herein described 30 polypeptide in an assay.

In one aspect, the activity is measured after 15 min. reaction, 30 min. reaction, 60 min. reaction, 90 min. reaction, 120 min. reaction or 180 min. reaction. Thus in one aspect, as an example the relative transgalactosylation activity is measured 35 minutes after addition of enzyme, such as 30 minutes after addition of enzyme, such as 60 minutes after addition of enzyme, such as 120 minutes after addition of enzyme or such as 180 minutes after addition of enzyme.

In the present context, the term "ratio of transgalactosylating activity;  $\beta$ -galactosidase activity" means ([Glucose]–[Galactose]/[Galactose]).

In the present context, the term "lactose has been transgalactosylated" means that a galactose molecule has been 45 covalently linked to the lactose molecule such as for example covalently linked to any of the free hydroxyl groups in the lactose molecule or as generated by internal transgalatosylation for example forming allolactose.

In the present context, the term "milk-based assay" means 50 an assay performed in milk, reconstituted milk or solutions containing main milk constituents such as for example lactose. In one embodiment, a milk-based assay is performed by preparing samples in 9% reconstituted milk from skimmed milk powder (such as e.g. Humana Milk Union, DE NW508 EG) giving a final concentration of lactose of 5% w/w. Enzymes are dosed based upon the LAU activity determined as described below giving the desired final concentration in LAU ml

A sample is taken prior to addition of enzyme and additional samples are taken at indicated time points and the enzymes are immediately inactivated by incubating at 95° C. for 10 minutes. Samples are diluted 1:10 and 2  $\mu$ L are applied onto activated (161° C. for 10 min) HPTLC silica gel 60 (Merck Cat#1.05541.0001) plates with a CAMAG Automatic 65 TLC Sampler 4. The TLC plates are eluted with an eluent containing (80) Acetonitril: (20) Ethylacetat: (50) 1-Pro-

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panol: (40) Water. Samples are visualised by heating (161° C. for 10 min) and allowed to cool down before soaking in 5% w/w H2SO4 in 99.9% w/w ethanol. Plates are developed with heating 161° C. for 3 min.

In one aspect, such an assay is as described in example 3. In the context of the present application, 1 lactase unit (1 LAU) is the amount of enzyme which releases 1 micromole glucose per minute in M-buffer at pH 6.5 and 37° C. with a lactase concentration of 4.75% w/v. M-buffer is prepared by dissolving 3.98 g  $\rm C_6H_5Na_3O_7$ -5  $\rm 2H_2O_4$ , 8.31 g citric acid, 0.9 g  $\rm K_2SO_4$ , 2.6 g  $\rm K_2HPO_4$ , 7.35 g  $\rm KH_2PO_4$ , 5.45 g KOH, 4.15 g, MgCl<sub>2</sub> 6H<sub>2</sub>O, 3.75 g CaCl2 2H<sub>2</sub>O and 1.4 g NaHCO<sub>3</sub> in 4 liter water, adding 12.5 ml 4N NaOH, adjusting to pH 6.5 using HCI, and adding water up to a total volume of 5 liter.

The activity in LAU of a specific lactase may be determined by direct measurement of glucose released from lactose under the conditions described above. The skilled person will know how to determine such activity. Alternatively, the activity may be determined by using the lactase activity assay described in Example 1 of the present application. Here, the activity is obtained by comparing to a standard curve with a lactase of known activity, and the activity of the unknown sample calculated from this. The lactase of known activity may e.g., be Lactozym obtained from Novozymes A/S, Denmark.

In the present context, the term "which polypeptide is freeze-dried" means that the polypeptide has been obtained by freeze-drying a liquid of the polypeptide at an appropriate pressure and for an appropriate period removing the water.

In the present context, the term "which polypeptide is in solution" relates to a polypeptide which is soluble in a solvent without precipitating out of solution. A solvent for this purpose includes any millieu in which the polypeptide may occur, such as an aqueous buffer or salt solution, a fermentation broth, or the cytoplasm of an expression host.

In the present context, the term "stabilizer" means any stabilizer for stabilizing the polypeptide e.g., a polyol such as e.g., glycerol or propylene glycol, a sugar or a sugar alcohol, lactic acid, boric add, or a boric acid derivative (e.g., an aromatic borate ester). In one aspect, the stabilizer is glycerol.

In the present context, the term "carbohydrate substrate" means an organic compound with the genera; formula  $C_m(H_2O)_n$ , that is, consisting only of carbon, hydrogen and oxygen, the last two in the 2:1 atom ratio such as a disaccharide.

In the present context, the term "disaccharide" is two monosaccharide units bound together by a covalent bond known as a olycosidic linkage formed via a dehydration reaction, resulting in the loss of a hydrogen atom from one monosaccharide and a hydroxyl group from the other. The formula of unmodified disaccharides is  $\rm C_{12}H_{22}O_{11}$ . In one aspect, the disaccharide is lactulose, trehalose, rhamnose, maltose, sucrose, lactose, or celloblose. In a further aspect, the disaccharide is lactose.

The term "isolated" means that the sequence is at least substantially free from at least one other component with which the sequence is naturally associated in nature and as found in nature. In one aspect, "isolated polypeptide" as used herein refers to a polypeptide which is at least 30% pure, at least 40% pure, at least 60% pure, at least 80% pure, at least 90% pure, and at least 95% pure, as determined by SDS-PAGE.

The term "substantially pure polypeptide" means herein a polypeptide preparation which contains at most 10%, preferably at most 8%, more preferably at most 6%, more preferably at most 4%, at most 3%, even more preferably at most 2%, most preferably at most

1%, and even most preferably at most 0.5% by weight of other polypeptide material with which it is natively associated. It is, therefore, preferred that the substantially pure polypeptide is at least 92% pure, preferably at least 94% pure, more preferably at least 95% pure, more preferably at least 96% pure, 5 more preferably at least 96% pure, more preferably at least 97% pure, more preferably at least 98% pure, even more preferably at least 99%, most preferably at least 99.5% pure, and even most preferably 100% pure by weight of the total polypeptide material present in the preparation. The polypep- 10 tides disclosed herein are preferably in a substantially pure form. In particular, it is preferred that the polypeptides are in "essentially pure form", i.e., that the polypeptide preparation is essentially free of other polypeptide material with which it is natively associated. This can be accomplished, for 15 example, by preparing the polypeptide by means of wellknown recombinant methods or by classical purification methods. Herein, the term "substantially pure polypeptide" is synonymous with the terms "isolated polypeptide" and "polypeptide in isolated form."

The term "purified" or "pure" means that a given component is present at a high level state—e.g. at least about 51% pure, or at least about 75%, or at least about 80%, or at least about 90% pure, or at least about 95% pure or at least about 98% pure. The component is desirably the predominant active 25 component present in a composition.

The term "microorganism" in relation to the present invention includes any microorganism that could comprise a nucleotide sequence according to the present invention or a nucleotide sequence encoding for a polypeptide having the specific properties as defined herein and/or products obtained therefrom.

In the present context, "microorganism" may include any bacterium or fungus being able to ferment a milk substrate.

The term "host cell"—in relation to the present invention 35 includes any cell that comprises either a nucleotide, sequence encoding a polypeptide having the specific properties as defined herein or an expression vector as described above and which is used in the production of a polypeptide having the specific properties as defined herein. In one aspect, the production is recombinant production.

The term "milk", in the context of the present invention, is to be understood as the lacteal secretion obtained from any mammal, such as cows, sheep, goats, buffaloes or camels.

In the present context, the term "milk-based substrate" 45 means any raw and/or processed milk material or a material derived from milk constituents. Useful milk-based substrates include, but are not limited to solutions/suspensions of any milk or milk like products comprising lactose, such as whole or low fat milk, skim milk, buttermilk, reconstituted milk 50 powder, condensed milk, solutions of dried milk, UHT milk, whey, whey permeate, acid whey, or cream. Preferably, the milk-based substrate is milk or an aqueous solution of skim milk, powder. The milk-based substrate may be more concentrated than raw milk. In one embodiment, the milk-based 55 substrate has a ratio of protein to lactose of at least 0.2, preferably at least 0.3, at east 0.4, at east 0.5, at least 0.5 or, most preferably, at least 0.7. The milk-based substrate may be homogenized and/or pasteurized according to methods known in the art.

"Homogenizing" as used herein means intensive mixing to obtain a soluble suspension or emulsion. It may be performed so as to break up the milk fat into smaller sizes so that it no longer separates from the milk. This may be accomplished by forcing the milk at high pressure through small orifices.

"Pasteurizing" as used herein means reducing or eliminating the presence of live organisms, such as microorganisms, 10

in the milk-based substrate. Preferably, pasteurization is attained by maintaining a specified temperature for a specified period of time. The specified temperature is usually attained by heating. The temperature and duration may be selected in order to kill or inactivate certain bacteria, such as harmful bacteria, and/or to inactivate enzymes in the milk. A rapid cooling step may follow.

A "food product" or "food composition" in the context of the present invention may be any comestible food or feed product suitable for consumption by an animal or human.

A "dairy product" in the context of the present invention may be any food product wherein one of the major constituents is milk-based. Preferable, the major constituent is milk-based. More preferably, the major constituent is a milk-based substrate which has been treated with an enzyme having transgalactosylating activity.

In the present context, "one of the major constituents" means a constituent having a dry matter which constitutes more than 20%, preferably more than 30% or more than 40% of the total dry matter of the dairy product, whereas "the major constituent" means a constituent having a dry matter which constitutes more than 50%, preferably more than 60% or more than 70% of the total dry matter of the dairy product.

A "fermented dairy product" in present context is to be understood as any dairy product wherein any type of fermentation forms part of the production process. Examples of fermented dairy products are products like yoghurt, buttermilk, creme fraiche, quark and fromage frail. A fermented dairy product may be produced by any method known in the art.

The term "fermentation" means the conversion of carbohydrates into alcohols or acids through the action of a microorganism such as a starter culture. In one aspect, fermentation comprises conversion of lactose to lactic acid.

In the present context the term "Pfam domains" means regions within a protein sequence that are identified as either Pfam-A or Pfam-B based on multiple sequence alignments and the presence of Hidden Markov Motifs ("The Pfam protein families database": R. D. Finn, J. Mistry, Tate, P. Coggill, A. Heger, J. E. Pollington, O. L. Gavin, P. Gunesekaran, G. Ceric, K. Forslund, L. Holm, E. L. Sonnhammer, S. R. Eddy, A. Bateman Nucleic Acids Research (2010) Database Issue 38:D211-222.). As examples of Pfam domains mention may be made of Glyco\_hydro2N (PF02837), Glyco\_hydro (PF00703), Glyco\_hydro 2C (PF02836) and Bacterial Ig-like domain (group 4) (PF07532).

As used herein "a position corresponding to position" means that an alignment as described herein is made between a particular query polypeptide and the reference polypeptide. The position corresponding to a specific position in the reference polypeptide is then identified as the corresponding amino acid in the alignment with the highest sequence identity.

In one aspect, a polypeptide having transgalactosylating activity comprising an amino acid sequence having

- a. at least 66% sequence identity to the amino acid sequence of SEQ ID NO: 1, and/or
- b. at least 66% sequence identity to the amino acid sequence of SEQ ID NO: 2, is provided.

In one aspect, a polypeptide, wherein the amino acid sequence comprises at least one or more amino acid residue(s) selected from the following groups:

 a. an amino acid residue selected from the group consisting of D/E/N/Q at a position corresponding to position 576 in SEQ ID NO: 1,

- b. an amino acid residue selected from the group consisting of D/E/N/Q at a position corresponding to position 588 in SEQ ID NO: 1,
- c. an amino acid residue selected from the group consisting of E/D/Q/N at a position corresponding to position 592 5 in SEQ ID NO: 1 and/or
- d. an amino acid residue selected from the group consisting of D/E/Q/N at a position corresponding to position 625 in SEQ ID NO: 1, is provided.

In one aspect, a polypeptide, wherein the amino acid 10 sequence comprises at least one or more amino acid residue(s) selected from the following groups:

- a. an amino acid residue selected from the group consisting of D/E/N/Q at a position corresponding to position 592 in SEQ ID NO: 2,
- b. an amino acid residue selected from the group consisting of D/E/N/Q at a position corresponding to position 604 in SEQ ID NO: 2,
- c. an amino acid residue selected from the group consisting of E/D/O/N at a position corresponding to position 608 20 in SEQ ID NO: 2 and/or
- d. an amino acid residue selected from the group consisting of D/E/Q/N at a position corresponding to position 641 in SEQ ID NO: 2, is provided.

It has been found that the amino acid at a position corre- 25 sponding to position 576, 588, 592 and 625 in SEQ ID NO:1 and the respective amino acids at a position corresponding to position 592, 604, 608 and 641 in SEQ ID NO:2 have an effect on the activity of the polypeptides disclosed herein.

In one aspect, disclosed herein is a polypeptide, wherein 30 the amino acid sequence comprises at least one or more acidic amino acid residue(s) such as D or E, in a position corresponding to position 576, 588, 592 and 625 in SEQ ID NO:1 or in a position corresponding to position 592, 604, 608 and 641 in SEQ ID NO:2.

In another aspect, the present invention relates to a polypeptide having a ratio of transgalactosylating activity:βgalactosidase activity of at least 1 as measured at a concentration of 6 LAU/ml in a milk-based assay at 37° C. and 5 w/w % lactose after 30 minutes reaction comprising an amino acid 40 sequence having

- a. at least 66% sequence identity to the amino acid sequence of SEQ ID NO: 1, and/or
- b. at least 66% sequence identity to the amino acid sequence of SEQ ID NO: 2, is provided.

In a further aspect, a polypeptide comprising an amino acid sequence having at least 60% sequence identity to the amino acid sequence of SEQ ID NO: 5, is provided. In a further aspect, a polypeptide comprising an amino acid sequence having at least 94% sequence identity to the amino acid 50 reaction. sequence of SEQ ID NO: 8, is provided. In a further aspect, the use of a polypeptide having transgalactosylating activity comprising an amino acid sequence having at least 60% sequence identity to the amino acid sequence of SEQ ID NO: 5 for producing galacto-oligosaccharides, is provided. In a 55 one or more of the following characteristics: further aspect, the use of a polypeptide having transgalactosylating activity comprising an amino acid sequence having at least 94% sequence identity to the amino acid sequence of SEQ ID NO: 8 for producing galacto-oligosaccharides, is provided.

In a further aspect, the use of a polypeptide having transgalactosylating activity comprising an amino acid sequence

- a. at least 60% sequence identity to the amino acid sequence of SEQ ID NO: 3,
- b. at least 60% sequence identity to the amino acid sequence of SEQ ID NO: 4,

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- c. at least 60% sequence identity to the amino acid sequence of SEO ID NO: 6, or
- d. at least 60% sequence identity to the amino acid sequence of SEQ ID NO: 7, for producing galacto-oligosaccharides, is provided.

In a further aspect, a polypeptide comprising an amino acid sequence having at least 66% sequence identity to the amino acid sequence of SEQ ID NO: 1, and at least 60% sequence identity to the amino acid sequence of SEQ ID NO: 5, is provided.

In a further aspect, a polypeptide comprising an amino acid sequence having at least 66% sequence identity to the amino acid sequence of SEQ ID NO: 2, and at least 94% sequence identity to the amino acid sequence of SEQ ID NO: 8, is provided.

In a further aspect, a polypeptide containing one or more Pfam domains selected from: Glyco\_hydro2N (PF02837), Glyco\_hydro (PF00703), Glyco\_hydro 2C (PF02836) and Bacterial Ig-like domain (group 4) (PF07532), is provided. In yet a further aspect, a polypeptide containing the Pfam Glyco\_hydro2N (PF02837), domains Glyco\_hydro (PF00703), Glyco\_hydro 2C (PF02836) and Bacterial Ig-like domain (group 4) (PF07532), is provided. In yet a further aspect, a polypeptide containing the Glyco\_hydro2N (PF02837), Glyco\_hydro (PF00703), and Glyco\_hydro 2C (PF02836) domains which domains constitutes the catalytic domain of the polypeptide, is provided.

In a further aspect, a polypeptide comprising an amino acid sequence and having a ratio of transgalactosylating activity: β-galactosidase activity of at least 1, at least 2.5, at least 3, at least 4, at least 5, at least 6, at least 7, at least 8, at least 9, at least 10, at least 11, or at least 12 as measured at a concentration of 6 LAU/ml in a milk-based assay at 37° C. and 5 w/w % lactose after 15 or 30 minutes reaction, is provided. In a further aspect, the polypeptide is derived from Ruminococcus hansenii or Ruminococcus lactaris.

In one aspect, the herein disclosed polypeptide(s) has a transgalactosylating activity such that more than 20%, more than 30%, more than 40%, up to 50% of the initial lactose is transgalactosylated as measured at a concentration of 6 LAU/ ml in a milk-based assay at 37° C. and 5 w/w % lactose after 30 minutes of reaction.

In a further aspect, the herein disclosed polypeptide(s) has a β-galactosidase activity such that less than 80%, less than 70%, less than 60%, less than 50%, less than 40%, less than 30%, less than 20% of the lactose has been hydrolysed as measured at a concentration of 6 LAU/ml in a milk-based assay at 37° C. and 5 w/w % lactose after 30 minutes of

In one aspect, the β-galactosidase activity and/or the transgalactosylating activity are measured at a concentration of 6 LAU/ml, 3 LAU/ml or 1 LAU/ml.

In a further aspect, the herein disclosed polypeptide(s) has

- a) a ratio of transgalactosylating activity:β-galactosidase activity of at least of at least 1, at least 2.5, at least 3, at least 4, at least 5, at least 6, at least 7, at east 8, at least 9, at least 10, at least 11, or at least 12 as measured at a concentration of 6 LAU/ml in a milk-based assay at 37° C. and 5 w/w % lactose after 30 minutes reaction, and/or
- b) has a transgalactosylating activity such that more than 20%, more than 30%, more than 40%, and up to 50% of the initial lactose has been transgalactosylated as measured at a concentration of 6 LAU/ml in a milk-based assay at 37° C. and 5 w/w % lactose after 30 minutes of reaction.

In a further aspect, a polypeptide comprising an amino acid sequence having at least 60% sequence identity to the amino acid sequence of SEQ ID NO: 5, is provided. In a further aspect, a polypeptide comprising an amino acid sequence having at least 94% sequence identity to the amino acid sequence of SEQ ID NO: 8, is provided. In yet a further aspect, a polypeptide comprising an amino acid sequence having at least 60% sequence identity to the amino acid sequence of SEQ ID NO: 3, is provided. In yet a further aspect, a polypeptide comprising an amino acid sequence having at least 60% sequence identity to the amino acid sequence of SEQ ID NO: 4, is provided. In yet a further aspect, a polypeptide comprising an amino acid sequence having at least 94% sequence identity to the amino acid sequence of SEQ ID NO: 6, is provided. In yet a further aspect, a polypeptide comprising an amino acid sequence having at least 94% sequence identity to the amino acid sequence of SEQ ID NO: 7, is provided.

Proteins are generally comprised of one or more functional 20 regions, commonly termed domains. The presence of different domains in varying combinations in different proteins gives rise to the diverse repertoire of proteins found in nature. One way of describing the domains are by the help of the Pfam database which is a large collection of protein domain 25 families as described in "The Pfam protein families database": R. D. Finn, 1 Mistry J. Tate, P. Coggill, A. Heger, J. E. Pollington, O. L. Gavin, P. Gunesekaran, G. Ceric, K. Forslund, L. Holm, E. L. Sonnhammer, S. R. Eddy, A. Bateman Nucleic Acids Research (2010) Database Issue 30 38:D211-222. Each family is represented by multiple sequence alignments and hidden Markov models (HMMs). In a further aspect, the present inventors have found that the herein provided polypeptide(s) contains one or more of the Pfam domains Glyco\_hydro2N (PF02837), Glyco\_hydro 35 (PF00703), Glyco\_hydro 2C (PF02836) and Bacterial Ig-like domain (group 4) (PF07532). In one aspect, the herein provided polypeptide(s) contains Glyco hydro2N (PF02837), Glyco\_hydro (PF00703), Glyco\_hydro 2C (PF02836) and Bacterial Ig-like domain (group 4) (PF07532).

In one aspect, the herein disclosed polypeptide(s) comprises an amino acid sequence having an amino acid residue selected from the group consisting of D, E, N and Q at a position corresponding to position 576 in SEQ ID NO: 1. In one aspect, the herein disclosed polypeptide(s) comprises an amino acid sequence having an amino acid residue selected from the group consisting of D, E and N at a position corresponding to position 576 in SEQ ID NO: 1. In one aspect, the herein disclosed polypeptide(s) comprises an amino acid sequence having an amino acid residue selected from the group consisting of D and E at a position corresponding to position 576 in SEQ ID NO: 1. In one aspect, the herein disclosed polypeptide(s) comprises an amino acid sequence the amino acid residue D at a position corresponding to position 576 in SEQ ID NO: 1.

In one aspect, the herein disclosed polypeptide(s) comprises an amino acid sequence having an amino acid residue selected from the group consisting of D, E, N and Q at a position corresponding to position 588 in SEQ ID NO: 1. In one aspect, the herein disclosed polypeptide(s) comprises an amino acid sequence having an amino acid residue selected from the group consisting of D, E and N at a position corresponding to position 588 in SEQ ID NO: 1. In one aspect, the herein disclosed polypeptide(s) comprises an amino acid sequence having an amino acid residue selected from the 65 group consisting of D and E at a position corresponding to position 588 in SEQ ID NO: 1. In one aspect, the herein

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disclosed polypeptide(s) comprises an amino acid sequence the amino acid residue D at a position corresponding to position 588 in SEQ ID NO: 1.

In one aspect, the herein disclosed polypeptide(s) comprises an amino acid sequence having an amino acid residue selected from the group consisting of D, E, N and Q at a position corresponding to position 592 in SEQ ID NO: 1. In one aspect, the herein disclosed polypeptide(s) comprises an amino acid sequence having an amino acid residue selected from the group consisting of D, E and Q at a position corresponding to position 592 in SEQ ID NO: 1. In one aspect, the herein disclosed polypeptide(s) comprises an amino acid sequence having an amino acid residue selected from the group consisting of D and E at a position corresponding to position 592 in SEQ ID NO: 1. In one aspect, the herein disclosed polypeptide(s) comprises an amino acid sequence the amino acid residue E at a position corresponding to position 592 in SEQ ID NO: 1.

In one aspect, the herein disclosed polypeptide(s) comprises an amino acid sequence having an amino acid residue selected from the group consisting of D, E, N and Q at a position corresponding to position 625 in SEQ ID NO: 1. In one aspect, the herein disclosed polypeptides) comprises an amino acid sequence having an amino acid residue selected from the group consisting of D, E and N at a position corresponding to position 625 in SEQ ID NO: 1. In one aspect, the herein disclosed polypeptide(s) comprises an amino acid sequence having an amino acid residue selected from the group consisting of D and E at a position corresponding to position 625 in SEQ ID NO: 1. In one aspect, the herein disclosed polypeptide(s) comprises an amino acid sequence the amino acid residue D at a position corresponding to position 625 in SEQ ID NO: 1.

In one aspect, the polypeptides have useful transgalactosylating activity over a range of pH of 4-9, such as 5-8, such as 5.5-7.5.

The present invention encompasses polypeptides having a certain degree of sequence identity or sequence homology with amino acid sequence(s) defined herein or with a polypeptide having the specific properties defined herein. The present invention encompasses, in particular, peptides having a degree of sequence identity with any one of SEQ ID NO: 1-8, defined below, or homologues thereof.

In one aspect, the homologous amino acid sequence and/or nucleotide sequence should provide and/or encode a polypeptide which retains the functional transgalactosylating activity and/or enhances the transgalactosylating activity compared to a polypeptide of SEQ ID NO: 1 or 2.

In the present context, a homologous sequence is taken to include an amino acid sequence which may be at least 66%, 70%, 75%, 78%, at least 80%, at least 85%, at least 90%, at least 95%, at least 96%, at least 97%, at least 98% or at least 99%, identical to the subject sequence. Typically, the homologues will comprise the same active sites etc. as the subject amino acid sequence. Although homology can also be considered in terms of similarity (i.e. amino acid residues having similar chemical properties/functions), in the context of the present invention it is preferred to express homology in terms of sequence identity.

Sequence identity comparisons can be conducted by eye, or more usually, with the aid of readily available sequence comparison programs. These commercially available computer programs use complex comparison algorithms to align two or more sequences that best reflect the evolutionary events that might have led to the difference(s) between the two or more sequences. Therefore, these algorithms operate with a scoring system rewarding alignment of identical or

similar amino acids and penalising the insertion of gaps, gap extensions and alignment of non-similar amino acids. The scoring system of the comparison algorithms include:

- i) assignment of a penalty score each time a gap is inserted (gap penalty score),
- ii) assignment of a penalty score each time an existing gap is extended with an extra position (extension penalty score).
- iii) assignment of high scores upon alignment of identical amino acids, and
- iv) assignment of variable scores upon alignment of nonidentical amino acids.

Most alignment programs allow the gap penalties to be modified. However, it is preferred to use the default values when using such software for sequence comparisons.

The scores given for alignment of non-identical amino acids are assigned according to a scoring matrix also called a substitution matrix. The scores provided in such substitution matrices are reflecting the fact that the likelihood of one 20 amino acid being substituted with another during evolution varies and depends on the physical/chemical nature of the amino acid to be substituted. For example, the likelihood of a polar amino acid being substituted with another polar amino acid is higher compared to being substituted with a hydro- 25 phobic amino acid. Therefore, the scoring matrix will assign the highest score for identical amino acids, lower score for non-identical but similar amino acids and even lower score for non-identical non-similar amino acids. The most frequently used scoring matrices are the PAM matrices (Dayhoff et al. (1978), Jones et al. (1992)), the BLOSUM matrices (Henikoff and Henikoff (1992)) and the Gonnet matrix (Gonnet et al. (1992)).

Suitable computer programs for carrying out such an alignment include, but are not limited to, Vector NTI (Invitrogen Corp.) and the ClustalV, ClustalW and ClustalW2 programs (Higgins D G & Sharp P M (1988), Higgins et al. (1992), Thompson et al. (1994), Larkin et al. (2007). A selection of different alignment tools is available from the ExPASy Proteomics server at www.expasy.org. Another example of software that can perform sequence alignment is BLAST (Basic Local Alignment Search Tool), which is available from the webpage of National Center for Biotechnology Information which can currently be found at www.ncbi.nlm.nih.gov/ and which was firstly described in Altschul et al. (1990) J. Mol. Biol. 215; 403-410.

Once the software has produced an alignment, it is possible to calculate % similarity and % sequence identity. The software typically does this as part of the sequence comparison and generates a numerical result.

In one embodiment, it is preferred to use the ClustalW software for performing sequence alignments. Preferably, alignment with ClustalW is performed with the following parameters for pairwise alignment:

Substitution matrix:	Gonnet 250	
Gap open penalty:	20	
Gap extension penalty:	0.2	
Gap end penalty:	None	

ClustalW2 is for example made available on the Internet by the European Bioinformatics Institute at the EMBL-EBI webpage www.ebl.ac.uk under tools—sequence analysis—ClustalW2. Currently, the exact address of the ClustalW2 tool is www.ebi.ac.uk/Tools/clustalW2.

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In another embodiment, it is preferred to use the program Align X in Vector NTI (Invitrogen) for performing sequence alignments. In one embodiment, Exp10 has been may be used with default settings:

Gap opening penalty: 10
Gap extension penalty: 0.05
Gapseparation penalty range: 8

In a particular embodiment, the percentage of identity of one amino acid sequence with, or to, another amino acid sequence is determined by the use of the score matrix: blosum62mt2 and the VectorNTI Pair wise alignment settings

Settings	K-tuple	1
	Number of best diagonals	5
	Window size	5
	Gap Penalty	3
	Gap opening Penalty	10
	Gap extension Penalty	0.1

Thus, the present invention also encompasses variants, homologues and derivatives of any amino acid sequence of a protein or polypeptide as defined herein, particularly those of SEQ ID NO: 1 or those of SEQ ID NO: 2, 3, 4, 5, 6, 7 or 8 defined below.

The sequences, particularly those of variants, homologues and derivatives of SEQ ID NO: 1, 2, 3, 4, 5, 6, 7 or 8 defined below, may also have deletions, insertions or substitutions of amino acid residues which produce a silent change and result in a functionally equivalent substance. Deliberate amino acid substitutions may be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues as long as the secondary binding activity of the substance is retained. For example, negatively charged amino acids include aspartic acid and glutamic add; positively charged amino acids include lysine and arginine; and amino acids with uncharged polar head groups having similar hydrophilicity values include leucine, isoleucine, valine, glycine, alanine, asparagine, glutamine, serine, threonine, phenylalanine, and tyrosine.

The present invention also encompasses conservative substitution (substitution and replacement are both used herein to mean the interchange of an existing amino acid residue, with an alternative residue) that may occur i.e. like-for-like substitution such as basic for basic, acidic for acidic, polar for polar etc. Non-conservative substitution may also occur i.e. from one class of residue to another or alternatively involving the inclusion of unnatural amino acids such as ornithine (hereinafter referred to as Z), diaminobutyric acid ornithine (hereinafter referred to as B), norleucine ornithine (hereinafter referred to as O), pyriylalanine, thienylalanine, naphthylalanine and phenylglycine.

Conservative substitutions that may be made are, for example within the groups of basic amino acids (Arginine, Lysine and Histidine), acidic amino acids (glutamic acid and aspartic acid), aliphatic amino acids (Alanine, Valine, Leucine, Isoleucine), polar amino acids (Glutamine, Asparagine, Serine, Threonine), aromatic amino acids (Phenylalanine, Tryptophan and Tyrosine), hydroxyl amino acids (Serine, Threonine), large amino acids (Phenylalanine and Tryptophan) and small amino acids (Glycine, Alanine).

In one embodiment, the polypeptide is a polypeptide having the sequence shown in SEQ ID NO: 1 or a polypeptide variant having at least at least 66%, at least 70%, at least 75%, at least 78%, at least 80%, at least 85%, at least 90%, at least 95%, at least 96%, at least 97%, at least 98% or at least 99% amino acid sequence identity therewith. In one embodiment,

the polypeptide is a polypeptide having the sequence shown in SEQ ID NO: 1 or a polypeptide variant having at least at least 70% amino acid sequence identity therewith. In one embodiment, the polypeptide is a polypeptide having the sequence shown in SEQ ID NO: 1 or a polypeptide variant having at least at least 75% amino acid sequence identity therewith. In one embodiment, the polypeptide is a polypeptide having the sequence shown SEQ ID NO: 1 or a polypeptide variant having at least at least 80% amino acid sequence identity therewith.

In one embodiment, the polypeptide is a polypeptide having the sequence shown in SEQ ID NO: 3 or a polypeptide variant having at least at least 60%, at least 65%, at least 70%, at least 75%, at least 78%, at least 80%, at least 85%, at least 90%, at least 95%, at least 96%, at least 97%, at least 98% or at least 99% amino acid sequence identity therewith.

In one embodiment, the polypeptide is a polypeptide having the sequence shown in SEQ ID NO: 4 or a polypeptide variant having at least 60%, at least 65%, at least 70%, at least 20 75%, at least 78%, at least 80%, at east 85%, at least 90%, at least 95%, at least 96%, at least 97%, at least 98% or at least 99% amino acid sequence identity therewith.

In one embodiment, the polypeptide is a polypeptide having the sequence shown in SEQ ID NO: 5 or a polypeptide 25 variant having at least 60%, at least 65%, at least 70%, at least 75%, at least 78%, at least 80%, at least 85%, at least 90%, at least 95%, at least 96%, at least 97%, at least 98% or at least 99% amino acid sequence identity therewith.

In one embodiment, the polypeptide is a polypeptide hav- 30 ing the sequence shown in SEQ ID NO: 2 or a polypeptide variant having at least 60%, at least 65%, at least 75%, at least 78%, at least 80%, at least 85%, at least 90%, at least 95%, at least 96%, at least 97%, at least 98% or at least 99% amino acid sequence identity therewith. In one embodiment, the 35 polypeptide is a polypeptide having the sequence shown in SEQ ID NO: 2 or a polypeptide variant having at least at least 70% amino acid sequence identity therewith. In one embodiment, the polypeptide is a polypeptide having the sequence shown in SEQ ID NO: 2 or a polypeptide variant having at 40 least at least 75% amino acid sequence identity therewith. In one embodiment, the polypeptide is a polypeptide having the sequence shown in SEQ ID NO: 2 or a polypeptide variant having at least at least 80% amino acid sequence identity therewith.

In one embodiment, the polypeptide is a polypeptide having the sequence shown in SEQ ID NO: 6 or a polypeptide variant having at Beast at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 94%, at least 95%, at least 96%, at least 97%, at least 50% or at least 99% amino acid sequence identity therewith.

In one embodiment, the polypeptide is a polypeptide variant having the sequence shown in SEQ ID NO: 7 or a polypeptide variant having at least at least 65%, at least 70%, at least 75%, at least 78%, at least 80% at least 85%, at least 90%, at 55 least 94%, at least 95%, at least 96%, at least 97%, at least 98% or at least 99% amino acid sequence identity therewith.

In one embodiment, the polypeptide is a polypeptide variant having the sequence shown in SEQ ID NO: 8 or a polypeptide variant having at least at east 65% at least 70%, at least 60%, at least 78%, at least 80%, at least 85%, at least 90%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98% or at least 99% amino acid sequence identity therewith.

In one aspect, the polypeptide sequence used in the present invention is in a purified form.

In one aspect, the polypeptide or protein for use in the present invention is in an isolated form.

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A "variant" or "variants" refers to either polypeptides or nucleic acids. The term "variant" may be used interchangeably with the term "mutant". Variants include insertions, substitutions, transversions, truncations, and/or inversions at one or more locations in the amino acid or nucleotide sequence, respectively. The phrases "variant polypeptide", "polypeptide variant", "polypeptide", "variant" and "variant enzyme" mean a polypeptide/protein that has an amino acid sequence that either has or comprises the amino acid sequence of or is modified compared to the amino acid sequence of SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, or 5. The variant polypeptides include a polypeptide having a certain percent, e.g., 60%, 65%, 66%, 68%, 70%, 72%, 74%, 76%, 78%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99%, of sequence identity with SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, or 8. As used herein, "parent enzymes," "parent sequence," "parent polypeptide" mean enzymes and polypeptides from which any of the variant polypeptides are based, e.g., SEQ ID NO: 1 or 2. A "parent nucleic acid" means a nucleic acid sequence encoding the parent polypeptide. The signal sequence of a "variant" may be the same or may differ from the signal sequence of the wild-type Ruminococcus lactaris or Blautia/ Ruminococcus hansenii or a Bacillus signal peptide or any signal sequence that will secrete the polypeptide. A variant may be expressed as a fusion protein containing a heterologous polypeptide. For example, the variant can comprise a signal peptide of another protein or a sequence designed to aid identification or purification of the expressed fusion protein, such as a His-Tag sequence.

To describe the various variants that are contemplated to be encompassed by the present disclosure, the following nomenclature will be adopted for ease of reference. Where the substitution includes a number and a letter, e.g., 592P, then this refers to {position according to the numbering system/substituted amino acid}. Accordingly, for example, the substitution of an amino acid to proline in position 592 is designated as 592P, Where the substitution includes a letter, a number, and a letter, e.g., D592P, then this refers to {original amino add/position according to the numbering system/substituted amino acid}. Accordingly, for example, the substitution of alanine with proline in position 592 is designated as A592P.

Where two or more substitutions are possible at a particular position, this will be designated by contiguous letters, which may optionally be separated by slash marks "/", e.g., G303ED or G303E/D.

Position(s) and substitutions are listed with reference to either SEQ ID NO: 1 or SEQ ID NO: 2. Equivalent positions in another sequence may be found by aligning this sequence with either SEQ ID NO: 1 or SEQ ID NO: 2 to find an alignment with the highest percent identity and thereafter determining which amino acid aligns to correspond with an amino acid of a specific position of either SEQ ID NO: 1 or SEQ ID NO: 2. Such alignment and use of one sequence as a first reference is simply a matter of routine for one of ordinary skill in the art.

"Variant nucleic acids" can include sequences that are complementary to sequences that are capable of hybridizing to the nucleotide sequences presented herein, in particular to SEQ ID NO:10-11. For example, a variant sequence is complementary to sequences capable of hybridizing under stringent conditions, e.g., 50° C. and 0.2×SSC (1×SSC=0.15 M NaCl, 0.015 M sodium citrate, pH 7.0), to the nucleotide sequences presented herein, in particular to SEQ ID NO: 10-11. More particularly, the term variant encompasses sequences that are complementary to sequences that are capable of hybridizing under highly stringent conditions, e.g., 65° C. and 0.1×SSC, to the nucleotide sequences presented

herein, in particular to SEQ ID NO: 10-11. The melting point (Tm) of a variant nucleic acid may be about 1, 2, or 3° C. lower than the Tm of the wild-type nucleic acid.

In one aspect, the present invention relates to isolated polypeptides having transgalactosylating activity as stated 5 above which are encoded by polynucleotides which hybridize under very low stringency conditions, preferably low stringency conditions, more preferably medium stringency conditions, more preferably medium-high stringency conditions, even more preferably high stringency conditions, and most 10 preferably very high stringency conditions with i) the nucleic acid sequence comprised in SEQ ID NO: 10 encoding the mature polypeptide of SEQ ID NO: 1; the cDNA sequence of i) or iii) the complementary strand of i) or ii) or with i) the nucleic acid sequence comprised in SEQ ID NO: 11 encoding 15 the mature polypeptide of SEQ ID NO: 2; ii) the cDNA sequence of i) or iii) the complementary strand of i) or ii); (J. Sambrook, E. F. Fritsch, and T. Maniatis, 1989, Molecular Cloning, A Laboratory Manual, 2d edition, Cold Spring Harbor, N.Y.). A subsequence of SEO ID NO: 10 or 11 contains 20 at least 100 contiguous nucleotides or preferably at least 200 continguous nucleotides. Moreover, the subsequence may encode a polypeptide fragment which has lactase activity.

The nucleotide sequence of SEQ ID NO: 10 or 11 or a subsequence thereof, as well as the amino acid sequence of 25 SEQ ID NO: 1 or 2 or a fragment thereof, may be used to design a nucleic acid probe to identify and done DNA encoding polypeptides having transgalactosylase activity from strains of different genera or species according to methods well known in the art. In particular, such probes can be used 30 for hybridization with the genomic or cDNA of the genus or species of interest, following standard Southern blotting procedures, in order to identify and isolate the corresponding gene therein. Such probes can be considerably shorter than the entire sequence, but should be at least 14, preferably at 35 least 25, more preferably at least 35, and most preferably at least 70 nucleotides in length. It is, however, preferred that the nucleic acid probe is at least 100 nucleotides in length. For example, the nucleic acid probe may be at least 200 nucleotides, preferably at least 300 nucleotides, more preferably at 40 least 400 nucleotides, or most preferably at least 500 nucleotides in length. Even longer probes may be used, e.g., nucleic acid probes which are at least 600 nucleotides, at least preferably at least 700 nucleotides, more preferably at least 800 nucleotides, or most preferably at least 900 nucleotides in 45 length. Both DNA and RNA probes can be used. The probes are typically labeled for detecting the corresponding gene (for example, with <sup>32</sup>P, <sup>3</sup>H, <sup>33</sup>S, biotin, or avidin). Such probes are encompassed by the present invention.

A genomic DNA library prepared from such other organisms may, therefore, be screened for DNA which hybridizes with the probes described above and which encodes a polypeptide having lactase activity. Genomic or other DNA from such other organisms may be separated by agarose or polyacrylamide gel electrophoresis, or other separation techniques. DNA from the libraries or the separated DNA may be transferred to and immobilized on nitrocellulose or other suitable carrier material. In order to identify a clone or DNA which is homologous with SEQ ID NO: 10 or 11 or a subsequence thereof, the carrier material is used in a Southern blot.

For purposes of the present invention, hybridization indicates that the nucleotide sequence hybridizes to a labelled nucleic: acid probe corresponding to the nucleotide sequence shown in SEQ ID NO: 10 or 11, its complementary strand, or a subsequence thereof, under very low to very high stringency conditions. Molecules to which the nucleic acid probe hybridizes under these conditions can be detected using X-ray film.

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In a preferred aspect, the nucleic acid probe is nucleotides 175 to 2011 or nucleotides 198 to 2040 of SEQ ID NO: 10 or SEQ ID NO: 11 respectively. In another preferred aspect, the nucleic acid probe is a polynucleotide sequence which encodes the polypeptide of SEQ ID NO: 1 or SEQ ID NO: 2, or a subsequence thereof. In another preferred aspect, the nucleic acid probe is SEQ ID NO: 10 or SEQ ID NO: 11. In another preferred aspect, the nucleic acid probe is the mature polypeptide coding region of SEQ ID NO: 10 or SEQ ID NO: 11.

For long probes of at least 100 nucleotides in length, very low to very high stringency conditions are defined as prehybridization and hybridization at 42° C. in 5×SSPE, 0.3% SDS, 200 g/ml sheared and denatured salmon sperm DNA, and either 25% formamide for very low and low stringencies, 35% formamide for medium and medium-high stringencies, or 50% formamide for high and very high stringencies, following standard Southern blotting procedures for 1.2 to 24 hours optimally.

For long probes of at least 100 nucleotides in length, the carrier material is finally washed three times each for 15 minutes using 2×SSC, 0.2% SDS preferably at least at 45° C. (very low stringency), more preferably at least at 50° C. (low stringency), more preferably at least at 55° C. (medium stringency), more preferably at least at 60° C. (medium-high stringency), even more preferably at least at 65° C. (high stringency), and most preferably at least at 70° C. (very high stringency).

In a particular embodiment, the wash is conducted using 0.2×SSC, 0.2% SDS preferably at least at 45° C. (very low stringency), more preferably at least at 50° C. (low stringency), more preferably at least at 55° C. (medium stringency), more preferably at least at 60° C. (medium-high stringency), even more preferably at least at 65° C. (high stringency), and most preferably at least at 70° C. (very high stringency). In another particular embodiment, the wash is conducted using 0.1×SSC, 0.2% SDS preferably at least at 45° C. (very low stringency), more preferably at least at 55° C. (medium stringency), more preferably at least at 55° C. (medium-high stringency), even more preferably at least at 65° C. (high stringency), and most preferably at least at 70° C. (very high stringency).

For short probes which are about 15 nucleotides to about 70 nucleotides in length, stringency conditions are defined as prehybridization, hybridization, and washing post-hybridization at about 5° C. to about 10° C. below the calculated T<sub>m</sub> using the calculation according to Bolton and McCarthy (1962, *Proceedings of the National Academy of Sciences USA* 48:1390) in 0.9 M NaCl, 0.09 M Tris-HCl pH 7.6, 6 mM EDTA, 0.5% NP-40, 1×Denhardt's solution, 1 mM sodium pyrophosphate, 1 mM sodium monobasic phosphate, 0.1 mM ATP, and 0.2 mg of yeast RNA per ml following standard Southern blotting procedures.

For short probes which are about 15 nucleotides to about 70 nucleotides in length, the carrier material is washed once in 6×SCC plus 0.1% SDS for 15 minutes and twice each for 15 minutes using 6×SSC at 5° C. to 10° C. below the calculated T

Under salt-containing hybridization conditions, the effective T<sub>m</sub> is what controls the degree of identity required between the probe and the filter bound DNA for successful hybridization. The effective T<sub>m</sub> may be determined using the formula below to determine the degree of identity required for
 two DNAs to hybridize under various stringency conditions.

(See www.ndsu.nodak.edu/instruct/mcclean/plsc731/dna/dna6.htm)

The G+C content of SEQ ID NO: 10 is 42% and the G+C content of SEQ ID NO: 11 is 44%. For medium stringency, the formamide is 35% and the Na<sup>+</sup> concentration for 5×SSPE 5 is 0.75 M.

Another relevant relationship is that a 1% mismatch of two DNAs lowers the  $T_m$  by 1.4° C. To determine the degree of identity required for two DNAs to hybridize under medium stringency conditions at 42° C., the following formula is used: 10

% Homology=100-[(Effective  $T_m$ -Hybridization Temperature)/1.4]

(See www.ndsu.nodak.edu/instruct/mcclean/plsc731/dna/dna6.htm)

The variant nucleic acids include a polynucleotide having a certain percent, e.g., 60%, 65%, 70%, 75%, 50%, 85%, 90%, 95%, or 99%, of sequence identity with the nucleic acid encoding SEQ ID NO: 1 or 2. In one aspect, a nucleic acid capable of encoding a polypeptide as disclosed herein, is 20 provided. In a further aspect, the herein disclosed nucleic acid has a nucleic acid sequence which is at least 60%, such as at least 65%, such as at least 75%, such as at least 80%, such as at least 85%, such as at least 90%, such as at least 95%, such as at least 95%, such as at least 95% identical SEQ ID NO: 10 25 or 11.

In one aspect, the polypeptides disclosed herein comprises an amino acid sequence having at least 66% sequence identity to the amino acid sequence of the mature polypeptide encoded by the nucleotide sequence encoding the transgalatosylase contained in DSM accession no: 20583. In one aspect, the polypeptides disclosed herein comprises an amino acid sequence having at least 66% sequence identity to the amino acid sequence of the mature polypeptide encoded by the nucleotide sequence encoding the transgalatosylase contained in ATCC accession no: 29176. All considerations and limitations relating to sequence identities and functionality discussed in terms of the SEQ ID NO: 1 or 2 apply mutatis mutandis to sequence identities and functionality of these polypeptides and nucleotides.

As used herein, the term "expression" refers to the process by which a polypeptide is produced based on the nucleic acid sequence of a gene. The process includes both transcription and translation.

As used herein, "polypeptide" is used interchangeably 45 with the terms "amino acid sequence", "enzyme", "peptide" and/or "protein". As used herein, "nucleotide sequence" or "nucleic add sequence" refers to an oligonucleotide sequence or polynucleotide sequence and variants, homologues, fragments and derivatives thereof. The nucleotide sequence may 50 be of genomic, synthetic or recombinant origin and may be double-stranded or single-stranded, whether representing the sense or anti-sense strand. As used herein, the term "nucleotide sequence" includes genomic DNA, cDNA, synthetic DNA, and RNA.

"Homologue" means an entity having a certain degree of identity or "homology" with the subject amino acid sequences and the subject nucleotide sequences. In one aspect, the subject amino acid sequence is SEQ ID NO: 1-8, and the subject nucleotide sequence preferably is SEQ ID 60 NO: 10-11

A "homologous sequence" includes a polynucleotide or a polypeptide having a certain percent, e.g., 80%, 85%, 90%, 95%, or 99%, of sequence identity with another sequence. Percent identity means that, when aligned, that percentage of 65 bases or amino acid residues are the same when comparing the two sequences. Amino acid sequences are not identical,

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where an amino acid is substituted, deleted, or added compared to the subject sequence. The percent sequence identity typically is measured with respect to the mature sequence of the subject protein, i.e., following removal of a signal sequence, for example, Typically, homologues will comprise the same active site residues as the subject amino acid sequence. Homologues also retain enzymatic activity, although the homologue may have different enzymatic properties than the wild-type

As used herein, "hybridization" includes the process by which a strand of nucleic acid joins with a complementary strand through base pairing, as well as the process of amplification as carried out in polymerase chain reaction (PCR) technologies. The variant nucleic acid may exist as single- or double-stranded DNA or RNA, an RNA/DNA heteroduplex or an RNA/DNA copolymer. As used herein, "copolymer" refers to a single nucleic acid strand that comprises both ribonucieotides and deoxyribonucleotides. The variant nucleic acid may be codon-optimized to further increase expression.

As used herein, a "synthetic" compound is produced by in vitro chemical or enzymatic synthesis. It includes, but is not limited to, variant nucleic acids made with optimal codon usage for host organisms, such as a yeast cell host or other expression hosts of choice.

As used herein, "transformed cell" includes cells, including both bacterial and fungal cells, which have been transformed by use of recombinant DNA techniques. Transformation typically occurs by insertion of one or more nucleotide sequences into a cell. The inserted nucleotide sequence may be a heterologous nucleotide sequence, i.e., is a sequence that is not natural to the cell that is to be transformed, such as a fusion protein.

As used herein, "operably linked" means that the described components are in a relationship permitting them to function in their intended manner. For example, a regulatory sequence operably linked to a coding sequence is ligated in such a way that expression of the coding sequence is achieved under condition compatible with the control sequences.

As used herein, the term "fragment" is defined as a polypeptide having one or more (several) amino acids deleted from the amino and/or carboxyl terminus for example of the polypeptide of SEQ ID NO:12 or 13; wherein the fragment has transgalactosylating activity.

In one aspect, the term "polypeptide fragment" is defined herein as a polypeptide having one or more (several) amino acids deleted from the amino and/or carboxyl terminus of the polypeptide of SEQ ID NO:1 or 2; wherein the fragment has transgalactosylating activity.

In one aspect, a fragment contains at least 500, 700, 900 or 1000 amino acid residues. In one aspect, a fragment contains at the most 1250, 1200, 1180, 1170, 1150 or 1125 amino acid residues.

In a further aspect, the length of the polypeptide disclosed 55 herein is 500 to 1250 amino acids. In a further aspect, the length of the polypeptide variant is 500 to 1200 amino acids. In a further aspect, the length of the polypeptide variant is 700 to 1170 amino acids. In a further aspect, the length of the polypeptide variant is 900 to 1180 amino acids. In a further aspect, the length of the polypeptide variant is 900 to 1150 amino acids. In a further aspect, the length of the polypeptide variant is 1000 to 1125 amino acids.

In one aspect, a plasmid comprising a nucleic acid as described herein, is provided.

In one aspect, an expression vector comprising a nucleic acid as described herein, or capable of expressing a polypeptide as described herein, is provided.

In a further aspect, a host cell comprising, preferably transformed with, a plasmid as described herein or an expression vector as described herein, is provided.

In a further aspect, a cell capable of expressing a polypeptide as described herein, is provided.

In one aspect, the host cell as described herein, or the cell as described herein is a bacterial, fungal or yeast cell.

In a further aspect, the host cell is selected from the group consisting of Ruminococcus, Bifidobacterium, Lactococcus, Lactobacillus, Streptococcus, Leuconostoc, Escherichia, 10 Bacillus, Streptomyces, Saccharomyces, Kluyveromyces, Candida, Torula, Torulopsis and Aspergillus.

In a further aspect, the host cell is selected from the group consisting of Ruminococcus hansenii, Bifidobacterium breve, Bifidobacterium longum, Bifidobacterium infantis, Bifido- 15 bacterium bifidum and Lactococcus lactis.

In a further aspect, a method of expressing a polypeptide as described herein comprises obtaining a host cell or a cell as described herein and expressing the polypeptide from the cell or host cell, and optionally purifying the polypeptide. Polypeptide Variants of SEQ ID NO: 1 or SEQ ID NO:2

In one aspect, a variant of SEQ ID NO:1 or 2 having a substitution at one or more positions which effects an altered property such as improved transgalactosylation, relative to SEQ ID NO: 1 or 2, is provided. Such variant polypeptides are 25 also referred to in this document for convenience as "variant polypeptide", "polypeptide variant" or "variant". In one aspect, the polypeptides as defined herein have an improved transgalactosylating activity as compared to the polypeptide of SEQ ID NO: 1 or 2. In another aspect, the polypeptides as 30 defined herein have an improved reaction velocity as compared to the polypeptide of SEQ ID No: 1 or 2.

In one aspect, the polypeptides and variants as defined herein exhibit enzyme activity. In one aspect, the polypeptides and the variant polypeptides described herein comprise 35 transgalactosylation activity.

In one aspect, the ratio of transgalactosylating activity:βgalactosidase activity is at least 2.5, such as at least 3, such as at least 4, such as at least 5, such as at least 6, such as at least 7, such as at least 8, such as at least 9, such as at least 10, such 40 as at least 11, or such as at least 12 after 30 min. reaction.

In one aspect, the polypeptides and the variants as defined herein are derivable from microbial sources, in particular from a filamentous fungus or yeast, or from a bacterium. The enzyme may, e.g., be derived from a strain of Agaricus, e.g. A. 45 bisporus; Ascovaginospora; Aspergillus, e.g. A. niger, A. awamori, A. foetidus, A. japonicus, A. oryzae; Candida; Chaetomium; Chaetotomastia; Dictyostelium, e.g. D. discoideum; Kluveromyces, e.g. K. fragilis, K. lactis; Mucor, e.g. M. javanicus, M. mucedo, M. subtilissimus; Neurospora, e.g. 50 N. crassa; Rhizomucor, e.g. R. pusillus; Rhizopus, e.g. R. arrhizus, R. japonicus, R. stolonifer; Sclerotinia, e.g. S. libertiana; Torula; Torulopsis; Trichophyton, e.g. T. rubrum; Whetzelinia, e.g. W. sclerotiorum; Bacillus, e.g. B. coagulans, B. circulans, B. megaterium, B. novalis, B. subtilis, B. 55 expression of the variant, when the variant is produced in a pumilus, B. stearothermophilus, B. thuringiensis; Bifidobacterium, e.g. B. Iongum, B. bifidum, B. animalis; Chryseobacterium; Citrobacter, e.g. C. freundii; Clostridium, e.g. C. perfringens; Diplodia, e.g. D. gossypina; Enterobacter, e.g. E. aerogenes, E. cloacae Edwardsiella, E. tarda; Erwinia, 60 e.g. E. herbicola; Escherichia, e.g. E. coli; Klebsiella, e.g. K. pneumoniae; Miriococcum; Myrothesium; Mucor; Neurospora, e.g. N. crassa; Proteus, e.g. P. vulgaris; Providencia, e.g. P. stuartii; Pycnoporus, e.g. Pycnoporus cinnabarinus, Pycnoporus sanguineus; Ruminococcus, e.g. R. torques; Sal- 65 monella, e.g. S. typhimurium; Serratia, e.g. S. liquefasciens, S. marcescens; Shigella, e.g. S. flexneri; Streptomyces, e.g. S.

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antibioticus, S. castaneoglobisporus, S. violeceoruber; Trametes; Trichoderma e.g. T. reesei, T. viride; Yersinia e.g. Y. enterocolitica.

An isolated and/or purified polypeptide comprising a polypeptide or a variant polypeptide as defined herein is provided. In one embodiment, the variant polypeptide is a mature form of the polypeptide (SEQ ID NO: 1 or 2). In one aspect, the variants include a C-terminal domain.

In one aspect, a variant polypeptide as defined herein includes variants wherein between one and about 25 amino acid residues have been added or deleted with respect to SEQ ID NO: 1 or SEQ ID NO: 2. In one aspect, the variant has the amino acid sequence of SEQ ID NO: 1 or SEQ ID NO: 2, wherein any number between one and about 25 amino acids have been substituted. In a further aspect, the variant has the amino acid sequence of SEQ ID NO: 1 or SEQ ID NO: 2, wherein any number between three and twelve amino acids has been substituted. In a further aspect, the variant has the amino acid sequence of SEQ ID NO: 1 or SEQ ID NO: 2, 20 wherein any number between five and nine amino acids has been substituted.

In one aspect, at least two, in another aspect at least three, and yet in another aspect at least five amino acids of SEQ ID NO: 1 or SEQ ID NO: 2 have been substituted.

In one aspect, the herein disclosed polypeptide(s) has the sequence of SEQ ID NO: 1 or 2.

In one aspect, the herein disclosed polypeptide(s) has the sequence of SEQ ID NO: 1 or 2, wherein the 10, such as 9, such as 8, such as 7, such as 6, such 5, such as 4, such as 3, such as 2, such as 1 amino acid in the N-terminal end are substituted and/or deleted.

In a further aspect, the length of the polypeptide variant is 500 to 1250 amino acids. In a further aspect, the length of the polypeptide variant is 500 to 1200 amino acids. In a further aspect, the length of the polypeptide variant is 700 to 1170 amino acids. In a further aspect, the length of the polypeptide variant is 900 to 1180 amino acids. In a further aspect, the length of the polypeptide variant is 900 to 1150 amino acids. In a further aspect, the length of the polypeptide variant is 1000 to 1125 amino acids.

Polypeptide Characterization

Enzymes and enzyme variants thereof can be characterized by their nucleic acid and primary polypeptide sequences, by three dimensional structural modeling, and/or by their specific activity. Additional characteristics of the polypeptide or polypeptide variants as defined herein include stability, pH range, oxidation stability, and thermostability, for example. Levels of expression and enzyme activity can be assessed using standard assays known to the artisan skilled in this field. In another aspect, variants demonstrate improved performance characteristics relative to the polypeptide with SEQ ID NO: 1 or 2, such as improved stability at high temperatures, e.g., 65-85° C.

An expression characteristic means an altered level of particular host cell. Expression generally relates to the amount of active variant that is recoverable from a fermentation broth using standard techniques known in this art over a given amount of time. Expression also can relate to the amount or rate of variant produced within the host cell or secreted by the host cell. Expression also can relate to the rate of translation of the mRNA encoding the variant polypeptide.

A nucleic acid complementary to a nucleic acid encoding any of the polypeptide variants as defined herein set forth herein is provided. Additionally, a nucleic acid capable of hybridizing to the complement is provided. In another embodiment, the sequence for use in the methods and com-

positions described here is a synthetic sequence. It includes, but is not limited to, sequences made with optimal codon usage for expression in host organisms, such as yeast.

The polypeptide variants as provided herein may be produced synthetically or through recombinant expression in a host cell, according to procedures well known in the art. In one aspect, the herein disclosed polypeptide(s) is recombinant polypeptide(s). The expressed polypeptide variant as defined herein optionally is isolated prior to use.

In another embodiment, the polypeptide variant as defined 10 herein is purified following expression. Methods of genetic modification and recombinant production of polypeptide variants are described, for example, in U.S. Pat. Nos. 7,371, 552, 7,166,453; 6,890,572; and 6,667,065; and U.S. Published Application Nos. 2007/0141693; 2007/0072270; 15 2007/0020731; 2007/0020727; 2006/0073583; 0019347; 2006/0018997; 2006/0008890; 2006/0008888; and 2005/0137111. The relevant teachings of these disclosures, including polypeptide-encoding polynucleotide sequences, primers, vectors, selection methods, host cells, purification 20 and reconstitution of expressed polypeptide variants, and characterization of polypeptide variants as defined herein, including useful buffers, pH ranges, Ca2+ concentrations, substrate concentrations and enzyme concentrations for enzymatic assays, are herein incorporated by reference.

In another embodiment, suitable host cells include a Gram positive bacterium selected from the group consisting of *Bacillus subtilis, B. licheniformis, B. lentus, B. brevis, B. stearothermophilus, B. alkalophilus, B. amyloliquefaciens, B. coagulans, B. circulans, B. lautus, B. thuringiensis, Streptomyces lividans, or S. murinus;* or a Gram negative bacterium, wherein said Gram negative bacterium is *Escherichia coil* or a *Pseudomonas* species. In one aspect, the host cell is a *B. subtilius* or *B. lichenformis*. In one embodiment, the host cell is *B. subtilis* and the expressed protein is engineered to comprise a *B. subtilis* signal sequence, as set forth in further detail below. In one aspect, the host cell expresses the polynucleotide as set out in the claims.

In some embodiments, a host cell is genetically engineered to express a polypeptide variant as defined herein with an 40 amino acid sequence having at least about 66%, 68%, 70%, 72%, 74%, 78%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 100% identity with the polypeptide of SEQ ID NO:1 or 2, in some embodiments, the polynucleotide encoding a polypeptide variant as defined herein 45 will have a nucleic acid sequence encoding the protein of SEO ID NO: 1 or a nucleic acid sequence having at least about 66%, 68%, 70%, 72%, 74%, 78%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99% or 100% sequence identity with a nucleic acid encoding the protein of SEQ ID NO: 1 or 2. In 50 one embodiment, the nucleic acid sequence has at least about 60%, 66%, 68%, 70%, 72%, 74%, 78%, 80% 85%, 90%, 95%, 96%, 97%, 98% or 99% sequence identity to the nucleic acid of SEQ ID NO: 10-11.

#### Vectors

In one aspect, the invention relates to a vector comprising a polynucleotide. In one aspect, a bacterial cell comprises the vector. In some embodiments, a DNA construct comprising a nucleic acid encoding a variant is transferred to a host cell in an expression vector that comprises regulatory sequences operably linked to an encoding sequence. The vector may be any vector that can be integrated into a fungal host cell genome and replicated when introduced into the host cell. The FGSC Catalogue of Strains, University of Missouri, lists suitable vectors. Additional examples of suitable expression 65 and/or integration vectors are provided in Sambrook et al., Molecular Cloning: A Laboratory Manual,  $3^{rd}$  ed., Cold

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Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (2001); Bennett et al., More Gene Manipulations in Fungi, Academic Press, San Diego (1991), pp. 396-428; and U.S. Pat. No. 5,874,276. Exemplary vectors include pFB6, pBR322, PUC18, pUC100 and pENTR/D, pDON<sup>TM</sup>201, pDONR<sup>TM</sup>221, pENTR<sup>TM</sup>, pGEW®3Z and pGEM®4Z. Exemplary for use in bacterial cells include pBR322 and pUC19, which permit replication in *E. coli*, and pE194, for example, which permits replication in *Bacillus*.

In some embodiments, a nucleic acid encoding a variant is operably linked to a suitable promoter, which allows transcription in the host cell. The promoter may be derived from genes encoding proteins either homologous or heterologous to the host cell. Suitable non-limiting examples of promoters include cbh1, cbh2, egl1, and egl2 promoters. In one embodiment, the promoter is one that is native to the host cell. For example, when *P. saccharophila* is the host, the promoter is a native *P. saccharophila* promoter. An "inducible promoter" is a promoter that is active under environmental or developmental regulation. In another embodiment, the promoter is one that is heterologous to the host cell.

In some embodiments, the coding sequence is operably linked to a DNA sequence encoding a signal sequence. A representative signal peptide is SEQ ID NO; 9 which is the native signal sequence of the *Bacillus subtilis* aprE precursor. In other embodiments, the DNA encoding the signal sequence is replaced with a nucleotide sequence encoding a signal sequence from other extra-cellular *Bacillus subtilis* pre-cursors. In one embodiment, the polynucleotide that encodes the signal sequence is immediately upstream and in-frame of the polynucleotide that encodes the polypeptide. The signal sequence may be selected from the same species as the host cell.

In additional embodiments, a signal sequence and a promoter sequence comprising a DNA construct or vector to be introduced into a fungal host cell are derived from the same source. In some embodiments, the expression vector also includes a termination sequence. In one embodiment, the termination sequence and the promoter sequence are derived from the same source. In another embodiment, the termination sequence is homologous to the host cell.

In some embodiments, an expression vector includes a selectable marker. Examples of suitable selectable markers include those that confer resistance to antimicrobial agents, e.g., hygromycin or phleomycin. Nutritional selective markers also are suitable and include amdS, argB, and pyr4. In one embodiment, the selective marker is the amdS gene, which encodes the enzyme acetamidase; it allows transformed cells to grow on acetamide as a nitrogen source. The use of an *A. nidulans* amdS gene as a selective marker is described in Kelley et al., *EMBO J.* 4: 475-479 (1985) and Penttila et al., *Gene* 61: 155-164 (198).

A suitable expression vector comprising a DNA construct with a polynucleotide encoding a variant may be any vector that is capable of replicating autonomously in a given host organism or integrating into the DNA of the host. In some embodiments, the expression vector is a plasmid. In some embodiments, two types of expression vectors for obtaining expression of genes are contemplated. The first expression vector comprises DNA sequences in which the promoter, coding region, and terminator all originate from the gene to be expressed. In some embodiments, gene truncation is obtained by deleting undesired DNA sequences to leave the domain to be expressed under control of its own transcriptional and translational regulatory sequences. The second type of expression vector is preassembled and contains sequences required for high-level transcription and a selectable marker.

In some embodiments, the coding region for a gene or part thereof is inserted into this general-purpose expression vector, such that it is under the transcriptional control of the expression construct promoter and terminator sequences. In some embodiments, genes or part thereof are inserted downstream of the strong cbh1 promoter.

Transformation, Expression and Culture of Host Cells

Introduction of a DNA construct or vector into a host cell includes techniques such as transformation; electroporation; nuclear microinjection; transduction; transfection, e.g., lipo- 10 fection mediated and DEAE-Dextrin mediated transfection; incubation with calcium phosphate DNA precipitate; high velocity bombardment with DNA-coated microprojectiles; and protoplast fusion. General transformation techniques are known in the art. See, e.g., Ausubel et al. (1987), supra, 15 chapter 9; Sambrook et al. (2001), supra; and Campbell at al., Curr. Genet. 16: 53-56 (1989). The expression of heterologous protein in Trichoderma is described, for example, in U.S. Pat. No. 6,022,725; U.S. Pat. No. 6,268,328; Harkki at al., Enzyme Microb. Technol. 13: 227-233 (1991); Harkki et 20 al., Bio Technol. 7: 596-603 (1989); EP 244,234; and EP 215, 594. In one embodiment, genetically stable transformants are constructed with vector systems whereby the nucleic acid encoding a variant is stably integrated into a host cell chromosome. Transformants are then purified by known tech- 25 niques.

In one non-limiting example, stable transformants including an amdS marker are distinguished from unstable transformants by their faster growth rate and the formation of circular colonies with a smooth, rather than ragged outline on 30 solid culture medium containing acetamide. Additionally, in some cases a further test of stability is conducted by growing the transformants on solid non-selective medium, e.g., a medium that lacks acetamide, harvesting spores from this culture medium and determining the percentage of these 35 spores that subsequently germinate and grow on selective medium containing acetamide. Other methods known in the art may be used to select transformants. Identification of Activity

To evaluate the expression of a variant in a host cell, assays 40 can measure the expressed protein, corresponding mRNA, or  $\beta$ -galactosidase activity. For example, suitable assays include Northern and Southern blotting, RT-PCR (reverse transcriptase polymerase chain reaction), and in situ hybridization, using an appropriately labeled hybridizing probe. Suitable assays also include measuring activity in a sample. Suitable assays of the activity of the variant include, but are not limited to, ONPG based assays or determining glucose in reaction mixtures such for example described in the examples herein.

Methods for Purifying Herein Disclosed Polypeptides

In general, a variant produced in cell culture is secreted into the medium and may be purified or isolated, e.g., by removing unwanted components from the cell culture medium. In some cases, a variant may be recovered from a cell lysate. In such 55 cases, the enzyme is purified from the cells in which it was produced using techniques routinely employed by those of skill in the art. Examples include, but are not limited to, affinity chromatography, ion-exchange chromatographic methods, including high resolution ion-exchange, hydropho- 60 bic interaction chromatography, two-phase partitioning, ethanol precipitation, reverse phase HPLC, chromatography on silica or on a cation-exchange resin, such as DEAF, chromatofocusing, SDS-PAGE, ammonium sulfate precipitation, and gel filtration using Sephadex G-75, for example. Depending on the intended use the herein disclosed polypeptide(s) may for example be either freeze-dried or prepared in a solu-

tion. In one aspect, the herein disclosed polypeptide(s) is freeze-dried form. In another aspect, the herein disclosed polypeptide(s) is in solution.

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Methods for Immobilising and Formulation of the Herein Disclosed Polypeptides

The polypeptide compositions may be prepared in accordance with methods known in the art and may be in the form of a liquid or a dry composition. For instance, the polypeptide composition may be in the form of a granulate or a microaranulate. The polypeptide to be included in the composition may be stabilized in accordance with methods known in the art.

The enzyme preparation, such as in the form of a food ingredient prepared according to the present invention, may be in the form of a solution or as a solid—depending on the use and/or the mode of application and/or the mode of administration. The solid form can be either as a dried enzyme powder or as a granulated enzyme.

Examples of dry enzyme formulations include spray dried products, mixer granulation products, layered products such as fluid bed granules, extruded or pelletized granules, prilled products, or lyophilised products.

The enzyme preparation, such as in the form of a food ingredient prepared according to the present invention, may be in the form of a solution or as a solid—depending on the use and/or the mode of application and/or the mode of administration. The solid form can be either as a dried enzyme powder or as a granulated enzyme.

In one aspect the invention provides an enzyme complex preparation comprising the enzyme complex according to the invention, an enzyme carrier and optionally a stabilizer and/or a preservative.

In yet a further aspect of the invention, the enzyme carrier is selected from the group consisting of glycerol or water.

In a further aspect, the preparation comprises a stabilizer. In one aspect, the stabilizer is selected from the group consisting of inorganic salts, polyols, sugars and combinations thereof. In one aspect, the stabilizer is an inorganic salt such as potassium chloride. In another aspect, the polyol is glycerol, propylene glycol, or sorbitol. In yet another aspect, the sugar is a small-molecule carbohydrate, in particular any of several sweet-tasting ones such as glucose, galactose, fructose and saccharose.

In yet at further aspect, the preparation comprises a preservative. In one aspect, the preservative is methyl paraben, propyl paraben, benzoate, sorbate or other food approved preservatives or a mixture thereof.

The method of the invention can be practiced with immobilized enzymes, e.g. an immobilized lactase or other galactooligosaccharide producing enzymes. The enzyme can be immobilized on any organic or inorganic support. Exemplary inorganic supports include alumina, celite, Dowex-1-chloride, glass beads and silica gel. Exemplary organic supports include DEAE-cellulose, alginate hydrogels or alginate beads or equivalents. In various aspects of the invention, immobilization of the lactase can be optimized by physical adsorption on to the inorganic support. Enzymes used to practice the invention can be immobilized in different media, including water, Tris-HCl buffer and phosphate buffered solution. The enzyme can be immobilized to any type of substrate, e.g. filters, fibers, columns, beads, colloids, gels, hydrogels, meshes and the like.

Use of the Herein Disclosed Polypeptides

In one aspect, a method for producing a dairy product by treating a milk-based substrate comprising lactose with a polypeptide as described herein is provided. In a further aspect, a method for producing a dairy product by treating a

milk-based substrate comprising lactose with a polypeptide having a relative transgalactosylation activity above 60%, such as above 70%, such as above 75% after 15 min. reaction, is provided. In one aspect, the relative transgalactosylation activity is above 3 after 30 min. reaction. In a further aspect, 5 the relative transgalactosylation activity is above 6 after 30 min. reaction. In yet a further aspect, the relative transgalactosylation activity is above 12 after 30 min. reaction. In one aspect, a method is provided, wherein the treatment with a polypeptide as described herein takes place at an optimal temperature for the activity of the enzyme. In a further aspect, the polypeptide is added to the milk-based substrate at a concentration of 0.01-1000 ppm. In yet a further aspect, the polypeptide is added to the milk-based substrate at a concentration of 0.1-100 ppm. In a further aspect, the polypeptide is 15 added to the milk-based substrate at a concentration of 1-10 ppm. In one aspect, a method further comprising fermenting a substrate such as a dairy product with a microorganism, is provided. In a further aspect, the dairy product is yogurt. In a further aspect, the treatment with the polypeptide and the 20 microorganism is performed essentially at the same time. In one aspect, the polypeptide and the microorganism are added to the milk-based substrate essentially at the same time.

In one aspect, a composition preferably a food composition, more preferably a dairy product comprising a cell or a 25 polypeptide as described herein, is provided.

In one aspect, a dairy product comprising a cell or a polypeptide as described herein, is provided. In one aspect, the polypeptide as defined herein is added in a concentration of 0.01-1000 ppm. In one aspect, a dairy product comprising 30 an inactivated polypeptide as defined herein, is provided. In one aspect, a dairy product comprising an inactivated polypeptide as defined herein in a concentration of 0.01-1000 ppm, is provided. In one aspect, a dairy product comprising GOS formed in situ by a polypeptide as defined herein, is 35 provided. In one aspect, a dairy product comprising a cell as defined herein, is provided.

A dairy product as described herein may be, e.g., skim milk, low fat milk, whole milk, cream, UHT milk, milk having an extended shelf life, a fermented milk product, cheese, 40 yoghurt, butter, dairy spread, butter milk, acidified milk drink, sour cream, whey based drink, ice cream, condensed milk, dulce de leche or a flavoured milk drink. A dairy product may be manufactured by any method known in the art.

A dairy product may additionally comprise non-milk components, e.g., vegetable components such as, e.g., vegetable oil, vegetable protein, and/or vegetable carbohydrates. Dairy products may also comprise further additives such as, e.g., enzymes, flavouring agents, microbial cultures such as probiotic cultures, salts, sweeteners, sugars, acids, fruit, fruit 50 juices, or any other component known in the art as a component of, or additive to, a dairy product.

In one embodiment of the invention, one or more milk components and/or milk fractions account for at east 50% (weight/weight), such as at least 70%, e.g., at least 80%, 55 preferably at least 90%, of the dairy product.

In one embodiment of the invention, one or more milk-based substrates having been treated with an enzyme as defined herein having transgalactosylating activity account for at least 50% (weight/weight), such as at least 70%, e.g. at 60 least 80%, preferably at least 90%, of the dairy product.

In one embodiment of the invention, the dairy product is a dairy product which is not enriched by addition of pre-produced galacto-oligosaccharides.

In one embodiment of the invention, the polypeptide- 65 treated milk-based substrate is not dried before being used as an ingredient in the dairy product.

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In one embodiment of the invention, the dairy product is ice cream. In the present context, ice cream may be any kind of ice cream such as full fat ice cream, low fat ice cream, or ice cream based on yoghurt or other fermented milk products. Ice cream may be manufactured by any method known in the art.

In one embodiment of the invention, the dairy product is milk or condensed milk.

In one embodiment of the invention, the dairy product is UHT milk, UHT milk in the context of the present invention is milk which has been subjected to a sterilization procedure which is intended to kill all microorganisms, including the bacterial spores. UHT (ultra high temperature) treatment may be, e.g., heat treatment for 30 seconds at 130° C., or heat treatment for one second at 145° C.

In one preferred embodiment of the invention, the dairy product is ESL milk. ESL milk in the present context is milk which has an extended shelf life due to microfiltration and/or heat treatment and which is able to stay fresh for at least 15 days, preferably for at least 20 days, on the store shelf at 2-5° C.

In another preferred embodiment of the invention, the dairy product is a fermented dairy product, e.g., yoghurt.

The microorganisms used for most fermented milk products are selected from the group of bacteria generally referred to as lactic acid bacteria. As used herein, the term "lactic add bacterium" designates a gram-positive, microaerophilic or anaerobic bacterium, which ferments sugars with the production of acids including lactic acid as the predominantly produced acid, acetic acid and propionic acid. The industrially most useful lactic, acid bacteria are found within the order "Lactobacillales" which includes Lactococcus spp., Streptococcus spp., Lactobacillus spp., Leuconostoc spp., Pseudoleuconostoc spp., Pediococcus spp., Brevibacterium spp., Enterococcus spp. and Propionibacterium spp. Additionally, lactic add producing bacteria belonging to the group of anaerobic bacteria, bifidobacteria, i.e. *Bifidobacterium* spp., which are frequently used as food cultures alone or in combination with lactic acid bacteria, are generally included in the group of lactic acid bacteria. Lactic acid bacteria are normally supplied to the dairy industry either as frozen or freeze-dried cultures for bulk starter propagation or as socalled "Direct Vat Set" (DVS) cultures, intended for direct inoculation into a fermentation vessel or vat for the production of a fermented dairy product. Such cultures are in general referred to as "starter cultures" or "starters".

Commonly used starter culture strains of lactic acid bacteria are generally divided into mesophilic organisms having optimum growth temperatures at about 30° C. and thermophilic organisms having optimum growth temperatures in the range of about 40 to about 45° C. Typical organisms belonging to the mesophilic group include Lactococcus lactis, Lactococcus lactis subsp. cremoris, Leuconostoc mesenteroides subsp. cremoris, Pseudoleuconostoc mesenteroldes subsp. cremoris, Pediococcus pentosaceus, Lactococcus lactis subsp. lactis biovar. diacetylactis, Lactobacillus casei subsp. casei and Lactobacillus paracasei subsp. paracasei. Thermophilic lactic acid bacterial species include as examples Streptococcus thermophilus, Enterococcus faecium, Lactobacillus delbrueckii subsp. lactis, Lactobacillus helveticus, Lactobacillus delbrueckii subsp. bulgaricus and Lactobacillus acidophilus, Also the anaerobic bacteria belonging to the genus Bifidobacterium including Bifidobacterium bifidum, Bifidobacterium animalis and Bifidobacterium longum are commonly used as dairy starter cultures and are generally included in the group of lactic acid bacteria. Additionally, species of *Propionibacteria* are used as dairy starter cultures, in particular in the manufacture of cheese. Additionally,

organisms belonging to the *Brevibacterium* genus are commonly used as food starter cultures.

Another group of microbial starter cultures are fungal cultures, including yeast cultures and cultures of filamentous fungi, which are particularly used in the manufacture of certain types of cheese and beverage. Examples of fungi include Penicillium roqueforti, Penicillium candidum, Geotrichum candidum, Torula kefir, Saccharomyces kefir and Saccharomyces cerevisiae.

In one embodiment of the present invention, the microorganism used for fermentation of the milk-based substrate is *Lactobacillus casei* or a mixture of *Streptococcus thermophilus* and *Lactobacillus delbrueckii* subsp. *bulgaricus*.

Fermentation processes to be used in a method of the present invention are well known and the person of skill in the 15 art will know how to select suitable process conditions, such as temperature, oxygen, amount and characteristics of microorganism's, additives such as e.g. carbohydrates, flavours, minerals, enzymes, and process time. Obviously, fermentation conditions are selected so as to support the achievement 20 of the present invention. As a result of fermentation, pH of the milk-based substrate will be lowered. The pH of a fermented dairy product of the invention may be, e.g., in the range 3.5-6, such as in the range 3.5-5, preferably in the range 3.8-4.8.

In one aspect, a method of using the polypeptides or using 25 any one or more of the above mentioned cell types for producing oligosaccharides, is provided. The oligosaccharides comprise, but are not limited to fructooligo-saccharides, galacto-oligosaccharides, isomalto-oligosaccharides, malto-oligosaccharides, lactosucrose and xylo-oligosaccharides.

In one embodiment of the invention, the oligosaccharides are produced by incubating the cell expressing the polypeptide in a medium that comprises a disaccharide substrate such as for example lactulose, trehalose, rhamnose, maltose, sucrose, lactose, or cellobiose. The incubation is carried out under conditions where oligosaccarides are produced. The cells may be part of a product selected from the group consisting of yoghurt, cheese, fermented milk products, dietary supplements, and probiotic comestible products. Alternatively, the oligosaccharides can be recovered and subsequently be added to the product of interest before or after its preparation.

In one aspect, the use of a herein disclosed cell for producing a product selected from the group consisting of yoghurt, cheese, fermented milk product, dietary supplement and probiotic comestible product, is provided.

In one aspect, the polypeptides described herein may be used to prepare cheese products and in methods for making the cheese products. Cheese products may e.g. be selected from the group consisting of cream cheese, cottage cheese, 50 and process cheese. By adding polypeptides the cheeses may contain significantly increased levels of galacto-oligosaccharides and reduced levels of lactose. In one aspect, the lactose levels in the final cheese product may be reduced by at least about 25 percent, preferably at least about 50 percent, and 55 more preferably at least about 75 percent. The polypeptides may be used to reduce Lactose in cheese products to less than about 1 gram per serving, an amount that can be tolerated by most lactose-intolerant individuals.

The cheese products provided herein are nutritionally-enhanced cheese products having increased soluble fiber content, reduced caloric content, excellent organoleptic properties, improved texture, and flavor. Further, the polypeptides described herein may reduce the glycemic index of the cheese products because GOS are more slowly absorbed than lactose 65 or its hydrolysis products. Finally, the polypeptides may reduce the cost of production of cheese products, particularly

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cream cheese products, because GOS surprisingly provide improved texture to the cream cheese product, thus permitting reduced use of stabilizers, or by allowing for increased moisture content without syneresis.

In a further aspect, a composition comprising a polypeptide as described herein and a carbohydrate substrate, is provided. In a further aspect, the carbohydrate substrate is a disaccharide. In a further aspect, the disaccharide is for example lactulose, trehalose, rhamnose, maltose, sucrose, lactose or cellobiose. In yet a further aspect, the carbohydrate substrate is lactose. The composition is prepared such that oligosaccarides are produced. The polypeptide as described herein may be part of a product selected from the group consisting of yoghurt, cheese, fermented milk products, dietary supplements, and probiotic comestible products. In one aspect, a composition comprising a polypeptide as described herein and a stabilizer, is provided. Examples of stabilizers is e.g., a polyol such as, e.g., glycerol or propylene glycol, a sugar or a sugar alcohol, lactic acid, boric add, or a boric acid derivative (e.g., an aromatic borate ester).

In one aspect, the use of a transgalactosylating polypeptide as disclosed herein or a cell as disclosed herein, for producing galacto-oligosaccharides, is provided. In one aspect, the use of a transgalactosylating polypeptide as disclosed herein or a cell as disclosed herein, for producing galacto-oligosaccharides to be part of a product selected from the group consisting of yoghurt, cheese, fermented dairy products, dietary supplements and probiotic comestible products, is provided. In one aspect, the product is yoghurt, cheese, or fermented dairy products. In one aspect, the use of a transgalactosylating polypeptide as disclosed herein or a cell as disclosed herein, for producing galacto-oligosaccharides to enhance the growth of Bifidobacterium, is provided. In one aspect, the use of a transgalactosylating polypeptide as disclosed herein or a cell as disclosed herein, for producing galacto-oligosaccharides to enhance the growth of Bifidobacterium in a mixed culture fermentation, is provided.

In one aspect, a process for producing a transgalactosylating polypeptide as disclosed herein, comprising culturing a cell as disclosed herein in a suitable culture medium under conditions permitting expression of said polypeptide, and recovering the resulting polypeptide from the culture, is provided. A process for producing galacto-oligosaccharides, comprising contacting of an polypeptide of as disclosed herein or a cell as disclosed herein with a milk-based solution comprising lactose, is provided.

Addition of oligosaccharides may enhance growth of either *Bifidobacterium* alone or of *Bifidobacterium* in a mixed culture.

The treatment of milk products with enzymes that converts lactose into monosaccharides or GOS have several advantages. First the products can be consumed by people with lactose intolerance that would otherwise exhibit symptoms such as flatulence and diarrhea. Secondly, dairy products treated with lactase will have a higher sweetness than similar untreated products due to the higher perceived sweetness of glucose and galactose compared to lactose. This effect is particularly interesting for applications such as yoghurt and ice-cream where high sweetness of the end product is desired and this allows for a net reduction of carbohydrates in the consumed product. Thirdly, in ice-cream production a phenomenon termed sandiness is often seen, where the lactose molecules crystallizes due to the relative low solubility of the lactose. When lactose is converted into monosaccharides or GOS the mouth feeling of the ice-cream is much improved over the non-treated products. The presence of a sandy feeling due to lactose crystallization can be eliminated and the raw

material costs can be decreased by replacement of skimmed milk powder by whey powder. The main effects of the enzymatic treatment were increased sweetness.

In one aspect, the transgalactosylating polypeptide(s) as disclosed herein may be used together with other enzymes 5 such as proteases such as chymosin or rennin, lipases such as phospholipases, amylases, transferases, and lactases. In one aspect, the transgalactosylating polypeptide(s) as disclosed herein may be used together with lactase. This may especially be useful when there is a desire to reduce residual lactose after 10 treatment with the transgalactosylating polypeptide(s) as disclosed herein especially at low lactose levels. A lactase in the context of the present invention is any glycoside hydrolase having the ability to hydrolyse the disaccharide lactose into constituent galactose and glucose monomers. The group of 15 lactases comprises but is not limited to enzymes assigned to subclass EC 3.2.1.108. Enzymes assigned to other subclasses, such as, e.g., EC 3.2.1.23, may also be lactases the context of the present invention. A lactase in the context of the invention may have other activities than the lactose hydrol- 20 ysing activity, such as for example a transgalactosylating activity. In the context of the invention, the lactose hydrolysing activity of the lactase may be referred to as its lactase activity or its beta-galactosidase activity. Enzymes having lactase activity to be used in a method of the present invention 25 may be of animal, of plant or of microbial origin. Preferred enzymes are obtained from microbial sources, in particular from a filamentous fungus or yeast, or from a bacterium. The enzyme may, e.g. be derived from a strain of Agaricus, e.g. A. bisporus; Ascovaginospora; Aspergillus, e.g. A. niger, A. 30 awamori, A. foetidus, A. japonicus, A. oryzae; Candida; Chaetomium; Chaetotomastia; Dictyostelium e.g. D. discoideum; Kluveromyces, e.g. K. fragilis, K. lactis; Mucor, e.g. M. javanicus, M. mucedo, M. subtilissimus; Neurospora, e.g. N. crassa; Rhizomucor, e.g. R. pusillus; Rhizopus, e.g. R. 35 arrhizus, R. japonicus, R. stolonifer; Sclerotinia, e.g. S. libertiana; Torula; Torulopsis; Trichophyton, e.g. T. rubrum; Whetzelinia, e.g. W. sclerotiorum; Bacillus, e.g. B. coagulans, B. circulans, B. megaterium, B. novalis, B. subtilis, B. pumilus, stearothermophilus, B. thuringiensis; Bifidobacte- 40 rium e.g. B. longum, B. bifidum, B. animalis; Chryseobacterium; Citrobacter, e.g. C. freundii; Clostridium, e.g. C. perfringens; Diplodia, e.g. D. gossypiria; Enterobacter, e.g. E. aerogenes, E. cloacae Edwardsiella, E. tarda; Erwinia, e.g. E. herbicola; Escherichia, e.g. E. coli; Klebsiella, e.g. K. 45 pneumoniae; Miriococcum; Myrothesium; Mucor; Neurospora, e.g. N. crassa; Proteus, e.g. P. vulgaris; Providencia, e.g. P. stuartii; Pycnoporus e.g. Pycnoporus cinnabarinus, Pycnoporus sanguineus; Ruminococcus, e.g. R. torques; Salmonella, e.g. S. typhimurium; Serratia, e.g. S. liquefasciens, 50 S. marcescens; Shigella, e.g. S. flexneri; Streptomyces, e.g. S. antibioticus, S. castaneoglobisporus, S. violeceoruber; Trametes; Trichoderma, e.g. T. reesei, T. viride; Yersinia, e.g. Y. enterocolitica. In one embodiment, the lactase is an intracellular component of microorganisms like Kluyveromyces 55 and Bacillus. Kluyveromyces, especially K. fragilis and K. lactis, and other fungi such as those of the genera Candida, Torula and Torulopsis, are a common source of fungal lactases, whereas B. coagulans and B circulans are well known sources for bacterial lactases. Several commercial lactase 60 preparations derived from these organisms are available such as Lactozym® (available from Novozymes, Denmark), HA-Lactase (available from Chr. Hansen, Denmark) and Maxilact® (available from DSM, the Netherlands), all from K. lactis. All these lactases are so called neutral lactases having a pH optimum between pH 6 and pH 8. When such lactases are used in the production of, e.g. low-lactose yoghurt, the

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enzyme treatment will either have to be done in a separate step before fermentation or rather high enzyme dosages have to be used, because their activity drop as the pH decreases during fermentation. Also, these lactases are not suitable for hydrolysis of lactose in milk performed at high temperature, which would in some cases be beneficial in order to keep the microbial count low and thus ensure good milk quality.

In one embodiment, the enzyme is a lactase from a bacterium, e.g. from the family is Bifidobacteriaceae, such as from the genus *Bifidobacterium* such as the lactase described in WO 2009/071539.

Further aspects according to the invention:

Aspect 1. An isolated polypeptide having transgalactosylating activity selected from the group consisting of:

- a. a polypeptide comprising an amino acid sequence having at least 66% sequence identity to the amino acid sequence of the mature polypeptide of SEQ ID NO: 1,
- a polypeptide comprising an amino acid sequence having at least 66% sequence identity to the amine acid sequence of the mature polypeptide of SEQ ID NO: 2,
- c. a polypeptide encoded by a polynucleotide that hybridizes under at least low stringency conditions with i) the nucleic acid sequence comprised in SEQ ID NO: 10 encoding the mature polypeptide of SEQ ID NO: 1; ii) the cDNA sequence of i) or iii) the complementary strand of i) or ii);
- d. a polypeptide encoded by a polynucleotide that hybridizes under at least low stringency conditions with i) the nucleic acid sequence comprised in SEQ ID NO: 11 encoding the mature polypeptide of SEQ ID NO: 2; ii) the cDNA sequence of i) or iii) the complementary strand of i) or ii);
- e. a polypeptide comprising a conservative substitution, deletion and/or insertion of one or more amino acids of SEO ID NO: 1.
- f. a polypeptide comprising a conservative substitution, deletion and/or insertion of one or more amino acids of SEO ID NO: 2.
- g. a polypeptide encoded by a polynucleotide comprising a nucleotide sequence having at least 70% identity to the nucleotide sequence encoding for the mature polypeptide of SEQ ID NO: 1 or the nucleotide sequence comprised in SEQ ID NO:10 encoding a mature polypeptide,
- h. a polypeptide encoded by a polynucleotide comprising a nucleotide sequence having at least 70% identity to the nucleotide sequence encoding for the mature polypeptide of SEQ ID NO: 2 or the nucleotide sequence comprised in SEQ ID NO:11 encoding a mature polypeptide,
- i. a polypeptide comprising an amino acid sequence having at least 66% sequence identity to the amino acid sequence of the mature polypeptide encoded by the nucleotide sequence encoding the transgalatosylase contained in DSM accession no: 20583, and
- j.a polypeptide comprising an amino acid sequence having at least 66% sequence identity to the amino acid sequence of the mature polypeptide encoded by the nucleotide sequence encoding the transgalatosylase, contained in ATCC accession no: 29176.

Aspect 2. The polypeptide according to aspect 1, wherein the polypeptide of above items a, c, e, g and i at the most has a length of 1806 amino acids and the polypeptide of above items b, d, f, h and j at the most has a length of 1767 amino acids

Aspect 3, A polypeptide having transgalactosylating activity selected from the group consisting of:

- a. a polypeptide comprising an amino acid sequence having at least 66% sequence identity to the amino acid sequence of the mature polypeptide of SEQ ID NO: 1,
- b. a polypeptide comprising an amino acid sequence having at least 66% sequence identity to the amino acid 5 sequence of the mature polypeptide of SEQ ID NO: 2,
- c. a polypeptide encoded by a polynucleotide that hybridizes under at least low stringency conditions with i) the nucleic acid sequence comprised in SEQ ID NO: 10 encoding the mature polypeptide of SEQ ID NO: 1; ii) 10 the cDNA sequence of i) the complementary strand of i) or ii):
- d. a polypeptide encoded by a polynucleotide that hybridizes under at least low stringency conditions with i) the nucleic acid sequence comprised in SEQ ID NO: 11 15 encoding the mature polypeptide of SEQ ID NO: 2; ii) the cDNA sequence of i) or iii) the complementary strand of i) car ii);
- e. a polypeptide comprising a conservative substitution, SEQ ID NO: 1, and
- f. a polypeptide comprising a conservative substitution, deletion and/or insertion of one or more amino acids of SEQ ID NO: 2.

Aspect 4. The polypeptide according to aspect 3, wherein 25 the polypeptide of above items a, c, and e at the most has a length of 1806 amino acids and the polypeptide of above items h, d, and f at the most has a length of 1767 amino acids.

Aspect 5. A polypeptide having transgalactosylating activity selected from the group consisting of:

- a. a polypeptide comprising an amino acid sequence having at least 66% sequence identity to the amino acid sequence of the mature polypeptide of SEQ ID NO: 1,
- b. a polypeptide encoded by a polynucleotide that hybridizes under at least low stringency conditions with i) the 35 nucleic acid sequence comprised in SEQ ID NO: 10 encoding the mature polypeptide of SEQ ID NO: 1; ii) the cDNA sequence of i) or iii) the complementary strand of i) or ii); and
- c. a polypeptide comprising a conservative substitution, 40 deletion and/or insertion of one or more amino acids of SEQ ID NO: 1.

Aspect 6. The polypeptide according to aspect 5, wherein the polypeptide of above items a, b, and c at the most has a length of 1806 amino acids.

Aspect 7. A polypeptide having transgalactosylating activity selected from the group consisting of:

- a. a polypeptide comprising an amino acid sequence having at least 66% sequence identity to the amino acid sequence of the mature polypeptide of SEQ ID NO: 2, 50
- b. a polypeptide encoded by a polynucleotide that hybridizes under at least low stringency conditions with i) the nucleic acid sequence comprised in SEQ ID NO: 11 encoding the mature polypeptide of SEQ ID NO: 2; ii) strand of i) or ii); and
- c. a polypeptide comprising a conservative substitution, deletion and/or insertion of one or more amino acids of SEQ ID NO: 2,

Aspect 8. The polypeptide according to aspect 7, wherein 60 the polypeptide of above items a, b and c at the most has a length of 1767 amino acids.

Aspect 9. The polypeptide according to any one of aspects 1-8 having a ratio of transgalactosylating activity:β-galactosidase activity of at least 1, at least 2.5, at least 3, at least 4, 65 at least 5, at least 6, at least 7, at least 8, at least 9, at least 10, at least 11, or at least 12.

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Aspect 10. The polypeptide according to any one of aspect 1-9, wherein the amino add sequence has at east 68%, 70%, 72%, 74%, 76%, 78%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99%, sequence identity to the mature amino add sequence of SEO ID NO: 1 or 2.

Aspect 11. The polypeptide according to aspects 1-10 containing the catalytic domain of glycosyl hydrolase class 2 (GH 2), preferably containing one or more Pfam domains selected from: Glyco\_hydro2N (PF02837), Glyco\_hydro (PF00703), Glyco\_hydro 2C (PF02536) and Bacterial Ig-like domain (group 4) (PF07532).

Aspect 12. The polypeptide according to any one of aspects 1-11 comprising or consisting of the amino acid sequence of

Aspect 13. The polypeptide according to any one of aspects 1-12 being a fragment of the mature polypeptide of SEQ ID

Aspect 14. The polypeptide according to any one of aspects deletion and/or insertion of one or more amino acids of 20 1-11 comprising or consisting of the amino acid sequence of SEQ ID NO: 2

> Aspect 15. The polypeptide according to any one of aspects 1-11 and 14 being a fragment of the mature polypeptide of SEO ID NO: 13.

> Aspect 16. A polypeptide having transgalactosylating activity comprising an amino acid sequence having

- a. at least 66% sequence identity to the amino acid sequence of SEQ ID NO: 1, and/or
- b. at least 66% sequence identity to the amino acid sequence of SEQ ID NO: 2,

Aspect 17. The polypeptide according to any one of aspects 1-16 comprising an amino acid sequence having at least 66% sequence identity to the amino acid sequence of SEQ ID NO:

Aspect 18. The polypeptide according to any one of aspects 1-17 provided that the polypeptide is not the beta-galactosidase from Ruminococcus hansenii having SEQ ID NO: 12.

Aspect 19. The polypeptide according to any one of aspects 146 comprising an amino acid sequence having at least 66% sequence identity to the amino acid sequence of SEQ ID NO:

Aspect 20. The polypeptide according to any one of aspects 1-16 and 19 provided that the polypeptide is not the betagalactosidase from Ruminococcus lactaris having SEQ ID NO: 13.

Aspect 21. The polypeptide according to any one of aspects 1-15 comprising an amino add sequence having at least 60% sequence identity to the amino acid sequence of SEQ ID NO:

Aspect 22. The polypeptide according to any one of aspects 1-21 comprising an amino add sequence having at least 94sequence identity to the amino acid sequence of SEQ ID

Aspect 23. The polypeptide according to any one of the the cDNA sequence of i) or iii) the complementary 55 aspects 1-22 containing one or more Pfam domains selected from: Glyco hydro2N (PF02837), Glyco hydro (PF00703), Glyco\_hydro 2C (PF02836) and Bacterial Ig-like domain (group 4) (PF07532).

Aspect 24. A polypeptide having a ratio of transgalactosylating activity:β-galactosidase activity of at least 1, at least 2.5, at least 3, at least 4, at least 5, at least 6, at least 7, at least 8, at least 9, at least 10, at least 11, or at least 12 as measured at a concentration of 6 LAU/ml in a milk-based assay at 37° C. and 5 w/w % lactose after 30 minutes reaction.

Aspect 25. The polypeptide according to any one of the aspects 1-24, which is derived from Ruminococcus hansenii or Ruminococcus lactaris.

Aspect 26. The polypeptide according to any one of the aspects 24-25, wherein the polypeptide comprises an amino acid sequence as defined in any one of aspects 1-23.

Aspect 27. The polypeptide according to any one of the aspects 1-26 having a ratio of transgalactosylating activity:  $\beta$ -galactosidase activity of at least 1, at least 2.5, at least 3, at least 4, at least 5, at least 6, at least 7, at least 8, at least 9, at least 10, at least 11, or at least 12 as measured at a concentration of 6 LAU/ml in a milk-based assay at 37° C. and 5 w/w % lactose after 30 minutes reaction.

Aspect 28. The polypeptide according to any one of the aspects 1-27, wherein the amino acid sequence comprises at least one or more amino acid residue(s) selected from the following groups:

- a. an amino acid residue selected from the group consisting of D/E/N/Q at a position corresponding to position 576 in SEQ ID NO: 1,
- b. an amino acid residue selected from the group consisting of D/E/N/Q at a position corresponding to position 583 in SEQ ID NO: 1,
- c. an amino acid residue selected from the group consisting of E/D/Q/N at a position corresponding to position 592 in SEQ ID NO: 1 and/or
- d. an amino acid residue selected from the group consisting of D/E/Q/N at a position corresponding to position 625 in SEQ ID NO: 1.

Aspect 29. The polypeptide according to any one of the aspects 1-28, wherein the amino acid sequence comprises at least one or more amino acid residue(s) selected from the 30 following groups:

- a. an amino acid residue selected from the group consisting of D/E/N/Q at a position corresponding to position 592 in SEQ ID NO: 2,
- b. an amino acid residue selected from the group consisting of D/E/N/Q at a position corresponding to position 604 in SEQ ID NO: 2,
- c. an amino acid residue selected from the group consisting of E/D/Q/N at a position corresponding to position 608 in SEQ ID NO: 2 and/or
- d. an amino acid residue selected from the group consisting of D/E/Q/N at a position corresponding to position 641 in SEQ ID NO: 2.

Aspect 30. The polypeptide according to any one of the aspects 1-29, wherein the percentage of identity of one amino acid sequence with, or to, another amino acid sequence is determined by the use of the score matrix: blosum62mt2 and the VectorNTI Pair wise alignment settings

Settings	K-tuple	1
	Number of best diagonals	5
	Window size	5
	Gap Penalty	3
	Gap opening Penalty	10
	Gap extension Penalty	0.1

Aspect 31. The polypeptide according to any one of the aspects 1-30, which polypeptide has a transgalactosylating activity such that more than 20%, more than 30%, more than 40%, and up to 50% of the initial lactose is transgalactosylated as measured at a concentration of 6 LAU/ml in a milk-based assay at 37° C. and 5 w/w % lactose after 30 minutes of reaction

Aspect 32. The polypeptide according to any one of the aspects 1-31, which polypeptide has a  $\beta$ -galactosidase activity such that less than 80%, less than 70%, less than 60%, less than 50%, less than 20% of

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the lactose has been hydrolysed as measured at a concentration of 6 LAU/ml in a milk-based assay at  $37^{\circ}$  C. and 5 w/w % lactose.

Aspect 33. The polypeptide according to any one of the aspects 1-32, wherein the activity is measured at a concentration of 3 LAU/ml or 1 LAU/ml.

Aspect 34. The polypeptide according to any one of the aspects 1-33, wherein the activity is measured 15 minutes after addition of polypeptide, 30 minutes after addition of polypeptide, 60 minutes after addition of polypeptide, 90 minutes after addition of polypeptide, 120 minutes after addition of polypeptide or 180 minutes after addition of polypeptide.

Aspect 35. The polypeptide according to any one of the aspects 1-34, wherein the amino acid sequence has at least 68%, 70%, 72%, 74%, 76%, 78%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99%, sequence identity to the amino acid sequence of SEQ ID NO: 1.

Aspect 36. The polypeptide according to any one of the aspects 1-35, wherein the amino acid sequence has at least 80% sequence identity to the amino acid sequence of SEQ ID NO: 1.

Aspect 37. The polypeptide according to any one of the aspects 1-34, wherein the amino acid sequence has at east 68%, 70%, 72%, 74%, 76%, 78%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99%, sequence identity to the amino acid sequence of SEQ ID NO: 2.

Aspect 38. The polypeptide according to any one of the aspects 1-34 and 37, wherein the amino acid sequence has at least 80% sequence identity to the amino acid sequence of SEQ ID NO: 2.

Aspect 39. The polypeptide according to any one of the aspects 1-38, wherein the amino acid sequence has at least 64%, 66%, 68%, 70% 72%, 74%, 76%, 78%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99%, sequence identity to the amino add sequence of SEQ ID NO:

Aspect 40. The polypeptide according to any one of the aspects 1-39, wherein the amino acid sequence has at least 95%, 96%, 97%, 98%, or 99%, sequence identity to the amino acid sequence of SEQ ID NO: 8.

Aspect 41. The polypeptide according to any one of the aspects 1-40, which polypeptide is a recombinant polypeptide.

Aspect 42. The polypeptide according to any one of the aspects 1-41, which polypeptide is freeze-dried.

Aspect 43. The polypeptide according to any one of the aspects 1-42, which polypeptide is in solution.

Aspect 44. The polypeptide according to any one of the aspects 1-43, which polypeptide is isolated.

Aspect 45. The polypeptide according to any one of the aspects 1-44, which polypeptide is purified.

Aspect 46. A polypeptide having the sequence of SEQ ID

Aspect 47. The polypeptide according to any one of the aspects 1-46 having one or more of the following characteristics:

- a) a ratio of transgalactosylating activity: β-galactosidase activity of at least 1, at least 2.5, at least 3, at least 4, at least 5, at least 6, at least 7, at least 8, at least 9, at least 10, at least 11, or at least 12 as measured at a concentration of 6 LAU/ml in a milk-based assay at 37° C. and 5 w/w % lactose after 30 minutes reaction, and/or
- b) has a transgalactosylating activity such that more than 20%, more than 30%, more than 40%, and up to 50% of the initial lactose has been transgalactosylated as mea-

sured at a concentration of 6 LAU/ml in a milk-based assay at  $37^{\circ}$  C. and 5 w/w % lactose after 30 minutes of reaction.

Aspect 48. A nucleic acid capable of encoding a polypeptide according to any one of the aspects 1-47.

Aspect 49. The nucleic acid according to aspect 48 having a nucleic acid sequence which is at least 60% identical to SEQ ID NO: 10 or 11.

Aspect 50. A plasmid comprising a nucleic acid according to any one of the aspects 48-49.

Aspect 51. An expression vector comprising a nucleic acid according to any one of the aspects 48-49, or capable of expressing a polypeptide according to any one of the aspects 1-47.

Aspect 52. A host cell comprising, preferably transformed with, a plasmid according to aspect 50 or an expression vector according to aspect 51.

Aspect 53. A cell capable of expressing a polypeptide according to any one of the aspects 1-47.

Aspect 54. The host cell according to aspect 52, or the cell according to aspect 53, which is a bacterial, fungal or yeast cell

Aspect 55. The cell according to aspect 53, wherein the cell is selected from the group consisting of *Ruminococcus*, *Bifi-* 25 dobacterium, *Lactococcus*, *Lactobacillus*, *Streptococcus*, *Leuconostoc*, *Escherichia*, *Bacillus*, *Streptomyces*, *Saccharomyces*, *Kluyverornyces*, *Candida*, *Torula*, *Torulopsis* and *Aspergillus*.

Aspect 56. The cell according to aspect 53, wherein the cell 30 is selected from the group consisting of *Ruminococcus hansenii*, *Ruminococcus lactaris*, *Bifidobacterium breve*, *Bifidobacterium longum*, *Bifidobacterium infantis*, *Bifidobacterium bifidum* and *Lactococcus lactis*.

Aspect 57. A method of expressing a polypeptide, the 35 aspects 53 and 55-56. method comprising obtaining a host cell or a cell according to any one of aspects 52-56 and expressing the polypeptide from the cell or host cell, and optionally purifying the polypeptide.

Aspect 57. A method of expressing a polypeptide, the 35 aspects 53 and 55-56. Aspect 78. A dairy defined in any one of the cell or host cell, and optionally purifying the polypeptide.

Aspect 58. A method of expressing a polypeptide, the method comprising obtaining a cell according aspect 53 and expressing the polypeptide from the cell, and optionally purifying the polypeptide.

defined in any 0.01-1000 ppm.

Aspect 80. A polypeptide as defined in any 0.01-1000 ppm.

Aspect 59. A composition comprising a polypeptide according to any one of aspects 1-47, preferably a food composition, more preferably a dairy product.

Aspect 60. A composition comprising a polypeptide as defined in any of aspects 1-47 and a stabilizer.

Aspect 61. A composition comprising a polypeptide as defined in any of aspects 1-47 and a carbohydrate substrate.

Aspect 62. The composition according to aspect 61, 50 wherein the carbohydrate substrate is a disaccharide.

Aspect 63. The composition according to aspect 62, wherein the disaccharide is lactase.

Aspect 64. A method for producing a dairy product by treating a milk-based substrate comprising lactose with a 55 polypeptide having a ratio of transgalactosylating activity:  $\beta$ -galactosidase activity of at least 1, at least 2.5, at least 3, at least 4, at least 5, at least 6, at least 7, at least 8, at least 9, at least 10, at least 11, or at least 12 as measured at a concentration of 6 LAU/ml in a milk-based assay at 37° C. and 5 w/w 60% lactose after 30 minutes reaction.

Aspect 65. A method for producing a food product by treating a substrate comprising lactose with a polypeptide as defined in any one of aspects 1-47.

Aspect 66. A method for producing a dairy product by 65 treating a milk-based substrate comprising lactose with a polypeptide according to any one of aspects 1-47.

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Aspect 67. The method according to any one of aspects 64-66 further treating the substrate with a hydrolysing beta-galactosidase.

Aspect 68. The method according to any one of aspects 64-67, wherein the polypeptide has a ratio of transgalactosylation activity as defined in aspect 64.

Aspect 69. The method according to any one of aspects 64-68, wherein the milk-based substrate is yoghurt, cheese, or fermented dairy products.

Aspect 70. The method according to any one of aspects 64-69 further comprising fermenting said substrate with a microorganism capable of fermenting said substrate.

Aspect 71. The method according to any one of aspects 64-70, wherein substrate such as the milk-based substrate is vogurt.

Aspect 72. The method according to any one of aspects 64-71, wherein the treatment with the polypeptide and the microorganism is performed essentially at the same time.

Aspect 73. The method according to any one of aspects 64-72, wherein the polypeptide and the microorganism are added to the milk-hared substrate essentially at the same time.

Aspect 74. The method according to any one of aspects 64-73, wherein the polypeptide Is derived from a microorganism of the genus *Ruminococcus*.

Aspect 75. Use of a cell of any one of aspects 53 and 55-56 for producing a product selected from the group consisting of yoghurt, cheese, fermented milk product, dietary supplement and probiotic comestible product.

Aspect 76. A food product, preferably a dairy product, comprising a transgalactosylating enzyme obtained from *Ruminococcus hansenii* or *Ruminococcus lactaris*, preferably as defined in item a-h in aspect 1, and more preferably a polypeptide as defined in any one of aspects 1-47.

Aspect 77. A dairy product comprising a cell of any one of aspects 53 and 55-56.

Aspect 78. A dairy product comprising a polypeptide as defined in any one of aspects 1-47.

Aspect 79. A dairy product comprising a polypeptide as defined in any one of aspects 1-47 in a concentration of 0.01-1000 ppm.

Aspect 80. A dairy product comprising an inactivated polypeptide as defined in any one of aspects 1-47.

Aspect 81. A dairy product comprising an inactivated polypeptide as defined in any one of aspects 1-47 in a con-45 centration of 0.01-1000 ppm.

Aspect 82. A dairy product comprising GOS formed in situ by a polypeptide as defined in any one of aspects 1-27.

Aspect 83. Use of a transgalactosylating polypeptide of any one of aspects 1-47 or a cell of any one of aspects 53 and 55-56, for producing galacto-oligosaccharides.

Aspect 84. Use of a transgalactosylating polypeptide of any one of aspects 1-47 or a cell of any one of aspects 53 and 55-56, for producing galacto-oligosaccharides to be part of a product selected from the group consisting of yoghurt, cheese, fermented dairy products, dietary supplements and probiotic comestible products.

Aspect 85. Use of a transgalactosylating polypeptide of any one of aspects 1-47 or a cell of any one of aspects 53 and 55-56, for producing galacto-oligosaccharides to enhance the growth of *Bifidobacterium*.

Aspect 86. Use of a transgalactosylating polypeptide of any one of aspects 1-47 or a cell of any one of aspects 53 and 55-56, for producing galacto-oligosaccharides to enhance the growth of *Bifidobacterium* in a mixed culture fermentation.

Aspect 87. A process for producing a transgalactosylating polypeptide of any one of aspects 1-47, comprising culturing a cell of any one of aspects 53 and 55-56 in a suitable culture

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medium under conditions permitting expression of said polypeptide, and recovering the resulting polypeptide from the culture.

Aspect 88. A process for producing galacto-oligosaccharides, comprising contacting of an polypeptide of any one of aspects 1-47 or a cell of any one of aspects 53 and 55-56 with a milk-based solution comprising lactose.

Aspect 89. A galacto-oligosaccharide or composition thereof obtained by treating a substrate comprising lactose with a polypeptide as defined in any one of aspects 1-47.

#### Example 1

# Production of Polypeptide

A synthetic *Ruminococcus hansenii* gene with codons optimised for expression in *Bacillus subtilis* was purchased from GeneART (Regensburg, Germany). The synthetic gene was cloned into the pBN *Bacillus subtilis* expression vector (FIG. 1) and transformed into the *Bacillus subtilis* strain BG6006. Transformants were restreaked twice onto LB plates containing 10 µg/mL Neomycin as selection.

A preculture was setup in LB media containing  $10 \,\mu\text{g/mL}$  Neomycin and cultivated for 7 hours at  $37^{\circ}$  C. and  $180 \,\text{rpm}$  25 shaking.  $500 \,\mu\text{l}$  of this preculture was used to inoculate  $50 \,\text{mL}$  Grant's modified medium containing  $10 \,\mu\text{g/mL}$  Neomycin at allowed to grow for  $48 \,\text{hours}$  at  $33^{\circ}$  C. and  $180 \,\text{rpm}$  shaking.

Cultures were harvested by centrifugation at 10.000×g and sterile filtered. The fermentation broths were up-concentrated using Sartorius Vivaspin20 MWCO 10,000 Dalton (Product code VS2002) at 4000 rpm in a tabletop centrifuge. The concentrate was stabilised with 20 w/w % glycerol.

Grant's modified media was prepared according to the following directions:

PART I (Autoclave)		
Soytone	10	g
Bring to	500	mL per liter
-	PART II	-
1M K <sub>2</sub> HPO <sub>4</sub>	3	mL
Glucose	75	g
Urea	3.6	g
Grant's 10X MOPS		mL
Bring to 400 mL per liter		

PART I (2 w/w % Soytone) was prepared, and treated in an autoclave for 20-25 mins.

PART II was prepared, and mixed with PART 1 and pH was adjusted to pH to 7.3 with HCl/NaOH.

The volume was brought to full volume and sterilized through  $0.22\text{-}\mathrm{um}$  PES filter.

10×MOPS Buffer was re tired according to the following 55 directions:

83.72 g	Tricine
7.17 g	KOH Pellets
12 g	NaCl
29.22 g	0.276M K2SO4
10 mL	0.528M MgCl2
10 mL Bring to 100 mL.	Grant's Micronutrients 100X

100× Micronutrients was prepared according to the following directions:

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Sodium Citrate • 2H2O	1.47 g
CaCl2•2H2O	1.47 g
FeSO4•7H2O	0.4 g
MnSO4•H2O	0.1 g
ZnSO4•H2O	0.1 g
CuCl2•2H2O	0.05 g
CoCl2•6H2O	0.1 g
Na2MoO4•2H2O	0.1 g
	-

The volume was reached with milliQ water. Sterilization was through 0.2 um PES filter. Protection from light was by wrapping in foil. Storing was at 4 C.

Determining the Hydrolysis Activity of the Enzyme Preparations  $^{15}$ 

Enzymatic activity of *Ruminococcus hansenii* (SEQ ID NO:1), *Ruminococcus lactaris* (SEQ ID NO:2) and *Bifidobacterium bifidum* BIF3d3 (truncated) (as described by Jørgensen et al. (2001), Appl. Microbiol. Biotechnol., 57: 647-652 and EP patent 1,283,876) were measured using the commercially available substrate 2-Nitrophenyl-β-D-Galactopyranoside (ONPG) (Sigma N1127).

1×ONPG buffer composition:

50 mM	Na-Citrate	
100 mM	NaPO4	
$2\mathrm{mM}$	CaCL2	
1 mM	MgCL2	
20 mM	ONPG	

Dilution series of above enzymes and Lactozym® (from Novozymes) as a standard control were made in 96 well microtiter plates. 75 µl of the dilutions were transferred to a new microtiter plate and mixed with 75 µl of 2× concentrated ONPG-buffer. Absorbance measurements were recorded at 450 nm on a Molecular Device SpectraMax controlled by the Softmax software package. The chamber was equilibrated to 37° C. and recordings were made every 15 seconds for 10 min in total. The ONP generation was measured and the Vmax of the reaction was determined. The Vmax for each enzyme preparation was compared to known concentrations of (3000 LAU/ml) Lactozym® and the activity in LAU/ml were calculated from the Lactozym® standard (see Table 1 below).

TABLE 1

Enzyme	LAU/ml
Lactozym ®	3000
Bifidobacterium bifidum BIF3d3 (truncated)*	105
Ruminococcus lactaris (SEQ ID NO: 2)	45
Ruminococcus hansenii (SEQ ID NO: 1)	42

# Example 2

#### Definition of GOS Producing Enzyme Unit

In the present application the relative transgalactosylation activity is defined as the difference between the amount of liberated glucose subtracted by the amount of liberated galactose divided by the amount of galactose generated in T-buffer at  $37^{\circ}~\rm C.$ 

Relative transgalactosylation activity =

Equation 1

[Glucose] – [Galactose]

T-buffer was prepared as follows:

50 mM	Na-citrate
100 mM	Na—PO4
2 mM	CaCl2
1 mM	MgCl2
5 w/w %	Lactose
pH 6.0	

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Measuring Galactose and Glucose by HPLC Chromatography

Galactose and glucose were analysed using a Dionex ICS3000 system consisting of ICS-3000 AS Autosampler, ICS-3000 ED Detector, ICS-3000 DC Chromatography Module and a DP Gradient pump (Dionex Corp, Sunnyvale, Calif., USA).

Galactose and glucose were separated using a CarboPac PA1 column 4 mm. with a CarboPac PA1 4 mm guard column (Dionex Corp, Sunnyvale, Calif., USA). The flow was 1 mL/min. The gradient was performed according to table 2, and the quantification was made with the use of external standards.

TABLE 2

Gradient program (w/w %) used for analysis of monosaccharides in samples		
Time (min)	Milli Q water	150 mM NaOH
0-12	90%-85%	10%-15%
12-25	85%-0%	15%-100%
25-30	0%	100%
30-32	0%-90%	100%-10%
32-34	90%	10%

The used eluents were water and 150 nM NaOH. 150 mM NaOH (eluent) was prepared by degassing 2 L Milli Q water for 10 min and adding 16 mL 50% w/w NaOH and degassing for another 5 min.

Calculation of Trangalactosylation Activity

The relative transgalactosylation activity was calculated according to equation 1 and the concentrations of glucose and galactose were measured by HPLC.

TABLE 3

Galactose concentration in %:									
Time/min	0	15	30	60	120	180			
Lactozym ®  Bifidobacterium bifidum	0	1.5 0.5	1.9 1	2.1 1	2.3 1	2.1 0.9			
BIF3d3 (truncated) Ruminococcus hansenii (SEQ ID NO: 1)	0	0.1	0.1	0.2	0.2	0.2			
Ruminococcus lactaris (SEQ ID NO: 2)	0	0.1	0.1	0.1	0.2	0.3			

TABLE 4

Glucose concentration in %:									
Time/min	0	15	30	60	120	180			
Lactozym ®	0	1.9	2.3	2.3	2.4	2.2			
Bifidobacterium bifidum BIF3d3 (truncated)	0	1.5	1.8	1.8	1.6	1.8			
Ruminococcus hansenii (SEO ID NO: 1)	0	0.7	1.3	1.2	1.3	1.3			

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TABLE 4-continued

Glucose concentration in %:									
Time/min	0	15	30	60	120	180			
Ruminococcus lactaris (SEQ ID NO: 2)	0	0.4	0.5	0.8	0.7	0.8			

#### TABLE

Ratio of transgalactosylating activity: β-galactosidase activity:								
Time/min	0	15	30	60	120	180		
Lactozym ®	nd	0.27	0.21	0.10	0.04	0.05		
Bifidobacterium bifidum BIF3d3 (truncated)	nd	2.00	0.80	0.80	0.60	1.00		
Ruminococcus hansenii (SEQ ID NO: 1)	nd	6.00	12.00	5.00	5.50	5.50		
Ruminococcus lactaris (SEQ ID NO: 2)	nd	3.00	4.00	7.00	2.50	1.67		

nd: Not determined for this timepoint.

TABLE 6

5	Relative transgalactosylation activity in %:								
	Time/min	0	15	30	60	120	180		
)	Lactozym ® Bifidobacterium bifidum BIF3d3 (truncated)	nd nd	21.05 66.67	17.39 44.44	8.70 44.44	4.17 37.50	4.55 50.00		
	Ruminococcus hansenii (SEQ ID NO: 1)	nd	85.71	92.31	83.33	84.62	84.62		
	Ruminococcus lactaris (SEQ ID NO: 2)	nd	75.00	80.00	87.50	71.43	62.50		

35 nd: Not determined for this timepoint.

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FIG. 2 displays the accumulation of glucose and galactose over time. As is clearly evident from FIG. 2 and the tables above, the *Ruminococcus hansenii* (SEQ ID NO:1) and *Ruminococcus lacteris* (SEQ ID NO:2) enzymes generate only between 10-20% of galactose relative to the *Bifidobacterium bifidum* BIF3d3 (truncated) enzyme. These finding suggest that both the *Ruminococcus hansenii* (SEQ ID NO:1) and *Ruminococcus lactaris* (SEQ ID NO:2) enzymes are able to exclude water from the active site more efficiently than Lactozym® and the *Bifidobacterium bifidum* BIF3d3 (truncated) enzyme.

# Example 3

# Assay in Milk

Samples were prepared in 9 w/w % reconstituted milk from skimmed milk powder (Humana Milk Union, DE NW508 EG) giving a final concentration of lactose of 5 w/w %. The enzymes were dosed based upon the LAU activity determined as described above at a final concentration of 6 LAU/ml. A sample was taken prior to addition of enzyme and additional samples were taken at indicated time points and the enzymes immediately inactivated by incubating at 95° C. for 10 minutes. Samples were diluted 1:10 and 2 µL were applied onto activated (161° C. for 10 min) HPTLC silica gel 60 (Merck Cat#1.05641.0001) plates with a CAMAG Automatic TLC Sampler 4. The TLC plates were eluted with an eluent containing (80) Acetonitril: (20) Ethylacetat: (50) 1-Propanol: (40) Water. Samples were visualised by heating (161° C. for

10 min) and allowed to cool down before soaking in 5 w/w % H2SO4 in 99.9% ethanol. Plates were developed with heating  $161^{\circ}$  C. for 3 min.

TABLE 7

	Compositi	on of standards:	
	Std A conc (w/w %)	Std B conc (w/w %)	Std C conc (w/w %)
Glucose	0.5	0.4	0.1
Lactose	0.3	0.2	0.5
Galactose	0.1	0.05	0.3

The sample number in FIG. 3 is as shown in below table. Sample Number:

1	Std A	8 Lactozym ® 120	15	B. bifidum 180	22	R. l
2	Std B	9 Lactozym ® 180	16	R. hansenii 0 min	23	R. l
3	Std C	10 B. bifidum 0 min	17	R. hansenii 15 min	24	R. i
4	Lactozym ® 0 min	11 B. bifidum 15 min	18	R. hansenii 30 min	25	R. i
5	Lactozym ® 15 min	12 B. bifidum 30 min	19	R. hansenii 60 min	26	R. l
6	Lactozym ® 30 min	13 B. bifidum 60 min	20	R. hansenii 120 min	27	R. l
7	Lactozym ® 60 min	14 B. hifidum 120	21	R. hansenii 180 min		

FIG. 3 shows the sugar composition of the milk at various time points during incubation. Whereas Lactozym® generates approximately equal amounts of glucose and galactose, the *Bifidobacterium bifidum* BIF3d3 (truncated), *Ruminococcus hansenii* (SEQ ID NO:1) and *Ruminococcus lactaris* (SEQ ID NO:2) enzymes all generate more glucose than galactose. These results are indicative of all these enzymes being able to perform transgalactosylation in reconstituted milk with an initial lactose concentration of 5 w/w %.

# Example 4

# Activity of Catalytic Core Mutants of Ruminococcus hansenii

Purification of the Enzyme from Crude Samples

Crude enzymes samples were obtained as described in example 1.

# Purification Method

Ion Exchange chromatography, Q HiTrap HP FF 5 ml

The column was prepared as described by the manufacturer and equilibrated in 20 mM Tris/HCl buffer, pH 8.0 (Buffer A).  $^{50}$ 

The sample (5 ml) was desalted in Buffer A and applied to the column at a flow rate of 4 ml/min. The column was washed with buffer A and the bound proteins were eluted with a linear gradient of 0-0.6 M NaCl in buffer A. During the entire run  $^{55}$  fractions of 4 ml were collected.

#### Activity Assay

90  $\mu L$  reaction buffer was mixed with 30  $\mu L$  of the indicated diluted sample (table 8) of enzyme in a 96-well Eppendorf twin tech PCR plate (Cat. 951020401) and incubated for 30 minutes at 42° C. in an Eppendorf Mastercycler Gradient PCR machine. The reaction was stopped by transferring the mixture to a Costar 9017 96-well plate containing 120  $\mu L$  10% Na-carbonate (Stop solution). The reactions were measured at 420 nm in a Molecular Devices Spectra Max 190 plate reader.

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	Protein, μg/ml	Diluted	OD420	Time/min	Activity/min, %
E592Q Frac.	62	10X	0.011	25	3
D625N Frac.	11	2X	0.168	25	50
D588N Frac. 14	6	1X	0.019	25	6
D576N Frac. 17	29	5X	0.270	5	81

TABLE 8-continued

lactaris 0 min lactaris 15 min lactaris 30 min lactaris 60 min lactaris 120 min lactaris 180 min

	Protein, μg/ml	Diluted	OD420	Time/min	Activity/min, %
WT Frac.	33	5.5x	0.335	5	100

Protein concentrations were adjusted to that of sample D588N by diluting With 50 mM Na-P buffer (ph 7.0) and activity was measured as described above.

Table 8 shows the protein concentration in the indicated fractions, fold of dilution to reach the concentration of D588N, the OD420 measurement, reaction time in minutes and relative activity per min to the *Ruminococcus hansenii* 45 wild type enzyme (WT).

FIG. 4 shows the results of the anion exchange chromatography of the above variants of the *Ruminococcus hansenii* (SEQ ID NO:1). The gel is a Nu-PAGE 4-12% acrylamide gel stained with coomassie brilliant blue staining.

Lane	Sample	
1	E592Q Crude	
2	Frac. 13 Eluate	
3	Frac. 14 Eluate	
4	D625N Crude	
5	Frac. 14 Eluate	
6	Frac. 15 Eluate	
7	D588N Crude	
8	Frac. 14 Eluate	
9	Frac. 17 Eluate	
13	D576N Crude	
14	Frac. 17 Eluate	
15	Frac. 18 Eluate	
16	WT Crude	
17	Frac. 17 Eluate	
18	Frac. 18 Eluate	

LIST OF SE	QUENCES 1 is a 1125	5 amino add	truncated			
fragment o	f SEQ ID NO: EPEQVAVKDY	: 12:		VSNAOTPTFD	DSKWRTLSLP	60
HDYSTEQEYS	QSLEAESGYL	PGGVGWYRKN	PTLGEEAKGK	RIRIDPDGVY	MNATVYVNGK	120
EVGTRPYGYT	PFSFDITDYI	SYDKENTIAV	KVDHQTPSSR	WYSGSGIYPS	VNLTTTNDVR	180
VDLNGI KVES	NNLEKEAGKT	VNTDVKTTVV	NGSKEAKNIT	ITRTVPKKGE	KPDKAIGTFT	240
TEAQEIGAGK	KTEISATVPV	KNPELWSVEN	PALYTIRTEV	KAGDKLLDSY	DTEYGFRYLN	300
FDTETGPQLN	GKNVKLKGVC	MRHDQGALGA	VANRRAI ERQ	VEILQEMGCN	SIRVTHNPAS	360
KDLIEVCNEK	GILVIEEVRD	GWHRAKNGNS	NDYSVWPEKA	IEEDNAILGK	EADMTWAEYD	420
LKAIMKRDQN	APSIIEWSLG	NEIQEGAGGS	GYAERADKLI	KWAKEADATK	TLTIGSNAVK	480
PGDWEQVSIG	DKLTKAGGTS	GTNYSDGASY	DKIHKSHPDW	KLYGSETASS	VNSRGIYSVT	540
GNQEATSDQQ	LTAYDNSRVN	WGALASQAWY	DVIQPDPVAG	KYVWTGFDYI	GEPTPWNGTD	600
PGAKGTWPSP	KNSYPGIIDT	AGPPKDSYYF	YQSQWNEEVN	TLRVLPAWNE	DVVKKNSDGT	660
VPVVVYSDAK	EVELPRTPAN	GGEKKSLGKK	TFKTETTKAG	YSYQVLENGK	KKHKDLYMEW	720
QVPYEAGTLE	AVAKDADGNV	IKDTWGESVV	KTTGEEAKLS	AKTDRNSIQA	DGKDLSYITV	780
DVTDKDGNIV	PDAANRVTHD	VQGAGKLVGV	DNGSSPDRDS	YKADNRKAFS	GKVLAIVQST	840
EKAGEITVTA	KADGLESSTV	KITTTPVKEE	PSERYVESYK	YSKSYYVKTG	TKPQLPKKIE	900
AQYSDRTKED	VAVKWDEISD	EQISKTGSFT	VEGTVGKRDI	TVRINMIDDV	AALLNYSGAT	960
QKGVKPQLPD	VRPAVLPDGT	VLAASFPVQW	DEKDADTFQK	PDEIVTVNGS	ADIFGKTIPV	1020
TASIRVQKED	IKIGSSVTNV	AKLSQNIQGS	DTLEAIKDGK	TEMSLNNDGG	PNESAWSNWD	1080
ASQKGTKEAE	LTFTFDTQQR	IGEIVIHPAK	DNNSIRHPDA	GTTEI		1125
	2 is 1150 a		runcated			
•	QAVRTESQTQ		NNYSSTAQRS	QNPNSNWKPY	FGDAGNAQGA	60
TFDDSKWEQV	SLPHDYSISQ	EYSKSMEAES	GYLGGGTGWY	RKNFTLSSDT	QGKPVRIDFD	120
GVYMNATVWV	NGHEVGTHPY	GYTSPSFDIT	DYVKYDGENT	IAVKVVNNTP	SSRWYSGSGI	180
YPDVDLTITD	DVHVDLNGTK	TVVPNLETEK	GSTVNTDVTA	TVANDSDAAK	SVAVRHTVFP	240
KDGSADQSIG	TVTTNAQSIA	AGATAEIQAT	VPVSNPELWS	VENPSLYTVR	TEVLVDGQVT	300
DTYDTEYGFR	YFNFDSNTGF	SLNGENMKLK	GVCMHHDQGS	LGAAAYDSAI	DRQVKILKEM	360
GCNSIRVTHN	PAAQDLIDAC	NEQGILVVEE	APDTWTRPKN	GNSNDYSVWF	NQTVASDNEI	420
LGATNGETWA	QFDLESMISR	DYNAPSVIMW	SLGNEVMEGI	SGGTDAEYEA	YATLKINWAY	480
DADNTRPMTI	GDNKLKANWQ	ISKTFARLLT	EKGGTVGFNY	ADGRVLDSYH	SSNSNWLLYG	540
SETASAINSR	GIYYRTTGGG	QTSDKQLTSY	DNSNVGWGAT	ASNAWYTVLT	RDFAAGEYVW	600
TGFDYLGEPT	PWNGTGSGAV	GSWPSPKNSY	FGIIDTAGFA	KDSYYFYQSQ	WNDDVTTLHV	660
LPAWNNNVVS	PDSSGNVPVV	VYSDAASVEL	FPQAKGSDTK	TSLGKKTHTQ	KTTDAGYTYQ	720
IYEGSDKNST	TDKNLYLTWN	VPYADGTVSA	VAYNSNGQKI	TDTVGQSSVT	TTGKASKLKA	780
SADHKKIAAD	GESLSYITVD	VTDANGNIVP	DAENPVKFTV	EGDGELVGVD	NGSSPDHDSY	840
QADNPKAFSG	KVLAIVKSTK	EAGTITVTAS	ADGLDSASVK	ITTTAVDNGS	TEKQIDSFKM	900
SPTYYVKVGS	TPELPEKIVT	RYTDGTSEEL	PVTWDAITED	QIAAAGSFQV	KGTVKGGYSV	960
AVNVNMIDEV	GGLLNYSTNT	AVGVAPVLPT	SPPAVLQDGT	VMDVTFPVTW	EDKAASAYDK	1020
AGTVTVNGTA	NVLGKEIAVT	ASVRVQEETI	TIGDSVSADA	LNLTQSVPAD	KQSDTLNAIK	1080
DGSTTISSNT	SGGANPTVWS	NYDYSQDGNT	TADIIPEYAT	EQRLGQTVTH	EARDSWSMRY	1140

-continued PDAGATEIYV 1150 SEQ ID NO: 3 is amino acid residues 559-649 of SEQ ID No: 1: VNWGALASQA WYDVIQRDFV AGEYVWTGFD YIGEPTPWNG TDPGAKGTWP SPKNSYFGII 60 DTAGPPKDSY YFYOSOWNEE VNTLAVLPAW N 91 SEQ ID NO: 4 is amino acid residues 579-649 of SEQ ID No: 1: AGEYVWTGFD YIGEPTPWNG TDPGAKGTWP SPKNSYFGII DTAGFPKDSY YFYQSQWNEE 60 VNTLHVLPAW N 71 SEQ ID NO: 5 is amino acid residues 579-636 of SEQ ID No: 1: AGEYVWTGFD YIGEPTPWNG TDPGAKGTWP SPKNSYFGII DTAGFPKDSY YFYQSQWN 58 SEQ ID NO: 6 is amino acid residues 575-665 of SEO ID No: 2: VGWGATASNA WYTVLTPDFA AGEYVWTGFD YLGEPTPWNG TGSGAVGSWP SPKNSYFGII 60 DTAGPAKDSY YPYOSOWNDD VTTLHVLPAW N 91 SEQ ID NO: 7 is amino acid residues 594-665 of SEO ID No: 2: AGEYVWTGFD YLGEPTPWNG TGSGAVGSWP SPKNSYFGYI DTAGFAKDSY YFYOSOWNDD 60 VTTLHVLPAW N 71 SEQ ID NO: 8 is amino acid residues 594-652 of SEQ ID No: 2: AGEYVWTGFD YLGEPTPWNG TGSGAVGSWP SPKNSYFGII DTAGPAKDSY YFYQSQWN 58 SEQ ID NO: 9 is a signal peptide from the pBN Bacillus subtilis expression vector: VRSKKLWISLLFALALIFTMAFGSTSSAQA SEQ ID NO: 10 is the nucleotide sequence encoding SEQ ID NO: 1 including sequence encoding the signal  $\tt gtgagaagcaaaaaattgtggatcagtttgctgtttgcttttagcgttaatctttacgatggcgttcggcagcaccatccagcgcgcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttagcgttag$ 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qqcaqcqqcatttataqatcaqtcaacctqacaacaacaatqatqtccatqtcqatctqaatqqcattaaaqtcqaaaqcaac acaattacacatacagtctttaaaaaaaggcgaaaaaccggataaagcgatcggcacatttacaacagaagcgcaagaaattggc gcaggcaaaaaaacagaaatcagcgcaacagtcacggttaaaaatccggaactgtggtcagttgaaaatccggcactgtataca attegeacagaagttaaageaggegataaaetgetggatagetatgatacagaatatggettteattatetgaaetttgataea gaaa caggettt cagetgaatggcaaaaaegttaaactgaaaggegtttgcatgcatcatgatcaaggegcaetttggegcagttgcaaatagaagagcaattgaacgccaagtcgaaattctgcaagaaatgggctgcaatagcattagagtcacacataatccggca agcaaagatctgattgaagtctgcaacgaaaaaggcattctggtcattgaagaagtttttgacggcttggcatagagcaaaaaat ggcaacagcaacgattatagcgtctggtttgaaaaagcgatcgaagaagataacgcgattctgggaaaagaagcggatatgact tgggcagaatatgatctgaaagcgattatgaaacgcgatcaaaatgcaccgagcattattgaatggtcactgggcaatgaaatt caagaaggcgcaggcggatcaggctatgcagaaagagcggataaactgatcaaatgggcgaaagaagcagacgcaacaaaaaca ctgacaattggcagcaatgtagttaaaagaggcgattgggaacaagttagcatcggcgataaacttacaaaagcaggcggaaca  ${ t tcaggcacaaattattcagatggcgcatcatatgataaaattcataaagaacatccggattggaaactgtatggctcagaaaca$ 

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gcatcatcagttaatagccgtggcatttattcagttacaggcaatcaagaagcaacaagcgatcaacaactgacagcgtatgat aatagcagagttaattggggagcactggcatcacaagcatggtatgatgttatccagagagattttgtcgcaggcgaatatgtt tggacaggetttgattatateggegaacegacacegtggaatggeacagateegggageaaaaggeacatggeegteacegaaa aacagctactttggcattatcgatacagcaggctttccgaaagattcatattatttttatcagagccagtggaatgaagaagtc tatqaaqcaqqcacacttqaaqcaqttqcqaaaqatqcaaaaqqcaacqtcattaaaqatacaqaaqqcaqaaqcqtcqttaaa acaacaggcgaagaagcaaaactgtcagcaaaaacggatcgcaatagcattcaagcagatggcaaagatctgtcatatattaca qtcqatqtcacaqataaaqatqqcaatattqttccqqatqcaqcaaataqaqtcacatttqatqtccaaqqcqcaqqaaaactq qttqqcqttqataatqqctcatcaccqqatcatqataqctataaaqcqqataaccqcaaaqcattttcaqqcaaaqttctqqca attgttcagtcaacagaaaaagcaggcgaaattacagttacagcaaaagcagatggcctggaatcaagcacagtcaaaatcaca acaacaccggttaaaqaaqaaccgagcgaaagatatgtcgaaagctataaatacagcaaaagctattatgtgaaaacaggcaca aaaccqcaactqccqaaaaaaattqaaqcqcaqtataqcqatcqcacaaaaqaqqatqttqcqqtcaaatqqqatqaaatctca gatgaacaaattagcaaaacaggcagctttacagttgaaggcacagttggcaaaagagatatcacagtcaacattaacatgatc gatgatgttgcagcactgctgaattattcaggcgcaacacaaaaaggcgttaaaaccgcaacttccggatgttagaccggcagtt ctgcctgatggcacagtcctggcagcatcatttccggttcagtgggatgaaaaagatgcggatacatttcagaaaccggatgaa aaaattggcagcagcgttacaaatgttgcaaaactgagccaaaatattcaaggcagcgatacactggaagcaatcaaagatggc gaagoogaactgacatttacatttgatacacagcaacgcattggogaaattgtcattcatttttgogaaagataacaactcaatc agatttccggatgctggcacaacagaaatctaa

SEQ ID NO: 11 is the nucleotide sequence encoding SEQ ID NO: 2 including sequence encoding the signal peptide:

ggcgtttcagttccggcactggcacaacaagcagttagaacagaaagccaaacacaaatgtcatcagatccggaactggtctat gtgaataactatagcagcacagcacaagaagccagaactttaacagcaactggaaattctacttcggagatgcgggaaatgca caaqqcqcaacatttqatqataqcaaatqqqaacaaqtttcactqccqcatqattattcaatcaqccaaqaatataqcaaatca atqqaaqcaqaatcaqqctatcttqqcqqaqqcacaqqctqqtatcqcaaaaattttacactqaqcaqcqatacacaaqqcaaa agagtccgcattgattttgatggcgtctatatgaatgcaacagtttgggttaatggccatgaagttggcacacatccgtatggc tatacaagetttagetttgatateacagattatgtgaaatatgatggegaaaacacaattgeagteaaagtegteaataataea ccqtcaaqcaqatqqtattcaqqctcaqqcatttataqaqatqtcqatqacaatcacaqatqttcatqttcatqttqatctqaacqq  $\verb|cacaa| a a gtt a caa cacega a cetgga a a cagaa a a a gg cage a cagte a a tacagat gtt a cage a a cagtt ge ga a t gatte cage a cagtt ge ga a t gatte cage a cage$ aaatqcacaatcaattqcaqcaqqcqcaacaqcaqaaattcaaqcaacqqttccqqtttcaaatcctqaactqtqqtcaqttqa aaatccgtcactgtatacagtcagaacagaagttctggtcgacggccaagtcacagatacatatgatacagaatatggctttcg qtcactqqqcaatqaaqttatqqaaqqcattaqcqqaqqcacaqatqcaqaatatqaaqcqacaqcqacqaaactqattaactq

#### -continued

ggcgtatgatgcggataatacacgtccgatgacaattggcgataacaaactgaaagcgaactggcagatctcaaaaacattttgc gagactgctgacagaaaaaggcggaacagtgggctttaattatgcagatggcagagttctggattcatatcatagcagcaatag caattggctgctgtatggctcagaaacagcatcagcgattaatagccgtggcatctattatagaacaacaggcggaggccaaac atcagataaacagctgacaagctatgataattcaaatgttggctggggagcaacagcatcaaatgcatggtatacagttctgac aagagatttttgcggcaggcgaatatgttttggacaggctttgattatctgggcgaaccgacaccgttggaatggcacaggctcagg cgcagttggctcatggccgtcaccgaaaaattcttattttggcattatcgatacagcaggcttcgcaaaagatagctattattt ttatcagagccagtggaatgatgttacaacactgcatgttcttccggcatggaataataatgtcgtcagcaaagattcatc aqqcaatqttccqqttqttqtttattcaqatqcqqcatcaqtcqaactqtttttttcaaqcaaaaqqcaqcqatacaaaaacaaq cctgggcaaaaaaacatttacacagaaaacaacagcaggctatacatatcagatctatgaaggctcagataaaaacagcac aacagacaaaaacctgtatctgacatggaatgttccgtatgcagatggaacagtttcagcagttgcgtataatagcaacggcca qaaaattacaqatacaqttqqccaqtcctcaqttacaacaacaqqcaaaqcqtcaaaactqaaaqcatcaqcqqatcataaaaa aattqcaqcqqatqqcqaatcactqtcatatatcacaqtcqatqtcacaqatqcqaatqqcaatattqttccqqatqcaqaaaa tegegteaaatttacagttgaaggegatggegaactggttggegttgataatggeteateaceggateatgatteatateaage qqataaccqcaaaqcattttcaqqcaaaqttctqqcaattqtqaaaaqcacaaaaqaaqctqqcacaattacaqttacaqcatc  ${\tt agcagatggcctggattcagcatcagtcaaaatcacaacaacagcagtcgataatggcagcacagaaaaacaaatcgatagctt}$ aaaatgagccgcacatattatgttaaagttggcagcacaccggaactgccggaaaaaattgtcacacgctatacagatggcaca tcagaagaactgcctgttacttgggatgcaattacagaagatcaaattgcagcagcaggctcatttcaagttaaaggcacagtc a a aggeggat at the agtheorem can extract gate the action of the aggregation of the aggggcgttgcaccggttctgccgacatcaagaccggcagttctgcaagatggcacagttatggatgttacatttccggtcacatgg gaagataaagcagcaagcgcatatgataaagcaggcacagtgacagtcaatggcacagcaaatgtttctgggcaaagaaattgca gttacagcgagcgttagagttcaggaagaaacaatcacaattggagattcagtttcagcggatgcactgaatctgacacaaagc gttccggcagataaacaaagcgatacactgaacgcaattaaagatggctcaacaacaattagctcaaatacaagcggaggcgca aatcogacagtttggagcaactatgactatagccaggatggcaatacgacagcggatatcattttttgaatatgcgacagaacaa agactgggccaaatcgttacacatttttgcgagagatagctggtcaatgagatatcctgatgcaggcgctacagaaatttatgtc

SEQ ID NO: 12 is a beta-galactosidase from Ruminococcus/Bleutia hansenil DSM 20583:

myffgrsaimmltvktrkelfmrkqrlarigaatlaavltvqgrngfsstvyakeepvrvkadsqtqmssepeqvavkdygsns artqnfdsdwkfnlgdvsnaqtptfddskwrtlslphdysieqeysqsleaesgylpggvgwyrknftlgeeakgkriridfdg vymnatvyvngkevgthpygytpfsfditdyisydkentiavkvdhqtpssrwysgsgiyrsvnitttndvhvdlngikvesnn lekeagktvntdvkttvvngskeeknitithtvfkkgekpdkaigtftteaqeigagkkteisetvpvknpelwsvenpalyti rtevkagdklldsydteygfhylnfdtetgfqlngknvklkgvcmhhdqgalgavanrraierqveilqemgcnsirvthnpas kdlievcnekgilvieevfdgwhrakngnsndysvwfekaieednailgkeadmtwaeydikaimkrdqnapsiiewslgneiq egaggsgyaeradklikwakeadatktltigsnavkrgdweqvsigdkltkaggtsatnysdgasydkihkehpdwklygseta ssvnsrgiysvtgnqeatsdqqltaydnsrvnwgalasqawydviqrdfvageyvwtgfdyigeptpwngtdpgakgtwpspkn syfgiidtagfpkdsyyfyqsqwneevntlhvlpawnedvvkknsdgtvpvvvysdakevelfftpanggekkslgkktfktet tkagysyqviengkkkhkdlymewqvpyeagtleavakdakqnvikdtegrsvvkttgeeeklsaktdrnsiqadgkdlsyitv dvtdkdgnivpdaanrvtfdvqgagklvgvdngsspdhdsykadnrkafsgkvlaivqstekageitvtakadgiesstvkitt tpvkeepseryvesykysksyyvktgtkpqlpkkeaqysdrtkedvavkwdeisdeqisktgsftvegtvgkrdityninmidd vaailnysgatqkgvkpqlpdvrpavlpdgtvlaasfpvqwdekdadtfqkpdeivtvngsadifgktipvtasirvqkedikl gssvtnvakisqniqgsdtleaikdgktemslnndggpnesawsnwdasqkgtkeaeltftfdtqqrigeivihfakdnnsirf

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pdagtteifvsetgkdgtwekvevkehlgeekdrvkayryeiapvtatyvkvkvvnanatdtgnrkpctaitevelkkaegsfk vnetaeleevkvgervlpnaayaldsysvpetdaavtaktkdnasltiipkhenvvrmilesedhkatknfavrmgeeetvlpd ddsrdypvekitatagseykpgtanegpvkyvldgkaethwhtnwsvsgegskpehrtvtlqlgndeeeapmidairymprsng angryteyeiqysidgdkwqtaatgeidkkqtgwmilgfeepvqakyvrfigthttsdqgndkhmavselrarvateapapsek ytitanvndktmgavtldsetgeyekgtkatitavpkegfafvnwtidgqevskenpyihtvetdatitanferievenegwvq tengweyyengqkvvgwkevsgkwyyfeenglmqtgwvfvnnhwyymdqwgamcigwvavdghwyymdqwgamctgwvsvnghw yhmdqwgemqtgwalvdsnwyylntdgsmaigwvavnghwyymdqwgamqtgwalvdsnwyylntdgsmaigwvavnghwyymdqwgamqtgwalvdsnwyylntdgsmaigwvavnghwyymdqwgamqtgwvlvgsdwyyintdgsmassqwidgyyvdasgkmk

SEQ ID NO: 13 is a glycosidase from Ruminococcus lactaris ATCC 29176: fddskweqvslphdysisqeysksmeaesgylgggtgwyrknftlssdtggkrvridfdgvymnatvwvnghevgthpygytsf sfditdyvkydqentiavkvvnntpssrwysqsqiyrdvdltitddvhvdlnqtkvttpnletekqstvntdvtatvandsdaa ksvavrhtvfpkdqsadqsiqtvttnaqsiaaqateelqatvpvsnpelwsvenpslytvrtevlvdqqvtdtydteyqfryfn  ${\tt fdsntgfslngenmklkgvcmhhdqgslgaaaydsaldrqvkllkemgcnslrvthnpaaqdlldacneqgilvveeafdtwtright of the constraints of the constra$ pkngnsndysvwfnqtvasdneilgatngetwaqfdlesmisrdynapsvlmwslgnevmegisggtdaeyeatatklinwaydsgraphicker. The statement of the state $adntrp \verb|mtigdnkl| kanwqisktfarlltekggtvgfnyadgrvldsyhssnsnwllygsetasalnsrglyyrttgggqtsdklender and the statement of the st$ qltsydnsnvgwgatasnawytvltrdfaageyvwtgfdylgeptpwngtgsgavgswpspknsyfglldtagfakdsyyfyqs  ${\tt qwnddvttlhvlpawnnnvvskdssgnvpvvvysdaasvelffqakgsdtktslgkktftqkttdagytyqlyegsdknsttdk}$ nlyltwnvpyadgtvsavaynsngqkitdtvgqssvtttgkasklkasadhkkiaadgeslsyitvdvtdangnivpdaenrvk ftvegdgelvgvdngsspdhdsyqednrkafsgkvlaivkstkeagtitvtasadgldsasvkitttavdngstekqidsfkms  $\verb|rtyyvkvgstpelpekivtrytdgtseelpvtwdaitedqiaaagsfqvkgtvkggysvavnvnmldevggllnystntavgva|$ pvlptsrpavlqdgtvmdvtfpvtwedkaasaydkagtvtvngtanvlgkelavtasvrvqeetitigdsysadalnltqsvpallederication and the statement of the ${\tt dkqsdtlnaikdgsttissntsgganptvwsnydysqdgnttadiifeyateqrlgqivthfardswsmrypdagateiyvspd}$  ${\tt gtnwakldttetigtesgnvkpytydfapvgatfvkfhltnstqatgttakactgiteielkvatgsrttnttaelqtitvngk}$  ${\tt evpqtaldskvyttpailaeie} a tak dnesvtvl payndviriive {\tt sedhqtrntyevrl neaeqttpdsdsrdypvskltase}$ gseqsttgvegpasnakdgdestlwhtrwsapaatsdqlwftyeleeetvldalryiprqgtadgqnngrvneyrvevstdgst wttvstgnwedsqdwklaeftepvaakyvrltgvhtygssaanvdkymsaaeirlrmaesktdiadaangvtvtapdsievaka daenpvmfdlsdivvkagdttirygvdyvisyenntdfgtaklvikgiykgasliyditvnpkkvdptdpdqpdkpdtpdngnd ngndnngqnnngtddgkkdpggsgvtdnknggnnsnngtaagnkanaaaktgdtanmllpmiaamlagtavvgtisirrrrr

SEQ ID NO: 14 is the nucleotide sequence encoding SEQ ID NO: 12 without the signal sequence:

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gaagtetgeaacgaaaaaggeattetggteattgaagaagtttttgaeggetggeatagageaaaaaatggeaacageaacgat tatagcgtctggtttgaaaaagcgatcgaagaagataacgcgattctgggaaaaagaagcggatatgacttgggcagaatatgat ctgaaagcgactatgaaacgcgatcaaaatgcaccgagcattattgaatggtcactgggcaatgaaattcaagaaggcgcaggc ggatcaggctatgcagaaagagcggataaactgatcaaatgggcgaaagaagcagacgcaacaaaaacactgacaattggcagc aatgcagttaaaagaggcgattgggaacaagttagcatcggcgataaacttacaaaagcaggcggaacatcaggcacaaattat tcagatggcgcatcatatgataaaattcataaagaacatccggattggaaactgtatggctcagaaacagcatcatcagttaat agccgtggcatttattcagttacaggcaatcaagaagcaacaagcgatcaacaactgacagcgtatgataatagcagagttaat tggggagcactggcatcacaagcatggtatgatgttatccagagagattttgtcgcaggcgaatatgtttggacaggctttgat tatatoggogaacogacacogtggaatggoacagatoogggagcaaaaggoacatggoogtcacogaaaaacagotactttggo attatcgatacagcaggctttccgaaagattcatattatttttatcagagccagtggaatgaagaagtcaatacactgcacgtt  $\tt cttccggcatggaatgaagatgtcgtcaaaaaaactcagatggcacagttccggttgttgtttattcagatgcgaaagaagtc$ tatagctatcaagttctggaaaacggcaaaaaaaacataaagatctgtatatggaatggcaagttccgtatgaagcaggcaca cttgaagcagttgcgaaagatgcaaaaggcaacgtcattaaagatacagaaggcagaagcgtcgttaaaacaacaggcgaagaa gcaaaactgtcagcaaaaacggatcgcaatagcattcaagcagatggcaaagatctgtcatatattacagtcgatgtcacagat a a a gat gg caat at tgt tccg gat gcag caa at a gag tcac at tt gat gt cca a gg cg cag gaa a act gg tt gg cgt tg at a at the companion of theggctcatcaccggatcatgatagctataaagcggataaccgcaaagcattttcaggcaaagttctggcaattgttcagtcaaca gaaaaagcaggcgaaattacagttacagcaaaagcagatggcctggaatcaagcacagtcaaaatcacaacaacaccggttaaa gaagaaccgagcgaaagatatgtcgaaagctataaatacagcaaaagctattatgtgaaaacaggcacaaaaccgcaactgccg aaaaaaattgaagcgcagtatagcgatcgcacaaaagaggatgttgcggtcaaatgggatgaaatctcagatgaacaaattagc aaaacaggcagctttacagttgaaggcacagttggcaaaagagatatcacagtcaacattaacatgatcgatgatgttgcagca ctqctqaattattcaqqcqcaacacaaaaaqqcqttaaaccqcaacttccqqatqttaqaccqqcaqttctqcctqatqqcaca gtcctggcagcatcatttccggttcagtgggatgaaaaagatgcggatacatttcagaaaccggatgaaattgttacagttaac qttacaaatqttqcaaaactqaqccaaaatattcaaqqcaqcqatacactqqaaqcaatcaaaqatqqcaaaacaqaaatqaqc ctqaataatqatqqcqqaccqaatqaatcaqcatqqtcaaattqqqatqcatcacaqaaaqqcacaaaaqaaqccqaactqaca ggcacaacagaaatctttgtttcagaaacaggcaaagatggcacatgggaaaaagttgaagtcaaagagcatattggcgaagaa aaagatcgcgtcaaagcatatcgctatgaaattgcaccggttacagcgacatatgttaaagttaaagtcgtcaatgcgaacgcg acagatacaggcaatagaaaaccgtgcacagcaattacagaagtcgaactgaaaaaagcagaaggcagctttaaagtcaacgaa acagcagaactggaagaagttaaagttggcgaacgtgttctgccgaatgcagcatatgcactggattcatattcagttccggaa  ${\tt acggatgcagcagttacagcaaaaacaaaagataatgcgagcctgacaatcctgccgaaacatgaaaatgtcgtcagaatgatt}$ ctggaaagcgaagaccataaagcgacgaaaaactttgcagttagaatgggcgaagaagaacagttctgccggatgatgattca agagattatccggtcgaaaaatcacagcaacaccaggctcagaatataaaccgggaacagcaaatgaaggaccggttaaatatgttctggatggcaaagcagaacacattggcatacaaattggtcagtttcaggcgaaggctcaaaaccggaacatagaacagtt  ${\tt acactgcaactgggcaatgatgaagaagaagcaccgatgattgacgcactgagatatatgccgagatcaaatggcgcaaatggc}$ agagttacggaatatgaaattcagtatagcctggatggcgataaatggcaaacagcagcaacaggcgaaatcgataaaaaacaa acaggctagatgatcctgggctttgaagaaccggttcaagcaaaatatgtccgctttattggcacacatacaacatcagatcag ggcaatgataaacatatggcagtttcagaactgaqagcaagagttgcaacagaagcaccggcaccgtcagagaagtatacaatt

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SEQ ID NO: 15 is the nucleotide sequence encoding SEQ ID NO: 13 without the signal sequence:

tatgtgaataactatagcagcacagcacaaagaagccagaactttaacagcaactggaaattctacttcggagatgcgggaaat gcacaaggcgcaacatttgatgatagcaaatgggaacaagtttcactgccgcatgattattcaatcagccaagaatatagcaaa tcaatggaagcagaatcaggctatcttggcggaggcacaggctggtatcgcaaaaattttacactgagcagcgatacacaaggc aaaagagtccgcattgattttgatggcgtctatatggaatgcaacagtttgggttaatggccatgaagttggcacacatccgtat ggctatacaagctttagctttgatatcacagattatgtgaaatatgatggcgaaaacacaattgcagtcaaaagtcgtcaataat  ${\tt acaccgtcaagcagatggtattcaggctcaggcatttatagagatgtcgatctgacaatcacagatgatgttcatgttgatctg}$ aacggcacaaaagttacaacaccgaacctggaaacagaaaaaggcagcacagtcaatacagatgttacagcaacagttgcgaat acaacaaatgcacaatcaattgcagcaggcgcaacagcagaaattcaagcaacggttccggtttcaaatcctgaactgtggtca gttgaaaatccgtcactgtatacagtcagaacagaagttctggtcgacggccaagtcacagatacatatgatacagaatatggc  $\verb|ctgggagcgaca| a t ggcgaaacat gggcaca | a t t t gatct ggaaag cat gatct cacar gatt a taat gcacc gt cag t catt$ atgtggtcactgggcaatgaagttatggaaggcattagcggaggcacagatgcagaatatgaagcgacagcgacgaaactgatt aactgggcgtatgatgcggataatacacgtccgatgacaattggcgataacaaactgaaagcgaactggcagatctcaaaaaca tttgcgagactgctgacagaaaaaggcggaacagtgggctttaattatgcagatggcagagttctggattcatatcatagcagc aatagcaattggctgctgtatggctcagaaacagcatcagcgattaatagccgtggcatctattatagaacaacaggcggaggc caaacatcagataaacagctgacaagctatgataattcaaatgttggctggggagcaacagcatcaaatgcatggtatacagtt ctgacaagagatttttgcggcaggcgaatatgttttggacaggctttgattatctgggcgaaccgacaccgtggaatggcacaggc ccaggcgcagttggctcatggccgtcaccgaaaaattcttattttggcattatcgatacagcaggcttcgcaaaagatagctat tatttttatcagagccagtggaatgatgatgttacaacactgcatgttcttccggcatggaataataatgtcgtcagcaaagat  ${\tt tcatcaggcaatgttccggttgttgtttattcagatgcggcatcagtcgaactgttttttcaagcaaaaggcagcgatacaaaa$ agcacaacagacaaaacctgtatctgacatggaatgttccgtatgcagatggaacagtttcagcagttgcgtataatagcaac ggccagaaaattacagatacagttggccagtcctcagttacaacaacaggcaaagcgtcaaaactgaaagcatcagcggatcat

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Trp Pro Ser Pro Lys Asn Ser Tyr Phe Gly Ile Ile Asp Thr Ala Gly
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- C	ontinued

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Tyr	Phe	Gly 675	Ile	Ile	Asp	Thr	Ala 680	Gly	Phe	Pro	Lys	Asp 685	Ser	Tyr	Tyr
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Pro 705	Ala	Trp	Asn	Glu	Asp 710	Val	Val	Lys	Lys	Asn 715	Ser	Asp	Gly	Thr	Val 720
Pro	Val	Val	Val	Tyr 725	Ser	Asp	Ala	Lys	Glu 730	Val	Glu	Leu	Phe	Phe 735	Thr
Pro	Ala	Asn	Gly 740	Gly	Glu	Lys	Lys	Ser 745	Leu	Gly	ГÀа	Lys	Thr 750	Phe	Lys
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ГÀв	Leu	Ser	Ala 820	Lys	Thr	Asp	Arg	Asn 825	Ser	Ile	Gln	Ala	Asp 830	Gly	Lys
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Thr	Val 1040		ı Ala	a Ala	. Ser	Phe 104		ro '	Val	Gln	Trp	Asp 1050		. Lys	Asp
Ala	Asp 1055		Phe	e Glr	Lys	Pro 106		ap (	Glu	Ile	Val	Thr 106		. Asn	Gly
Ser	Ala 1070		) Il∈	Ph∈	e Gly	Lys 107		hr :	Ile	Pro	Val	Thr 108		Ser	Ile
Arg	Val 1085		Lys	g Glu	ı Asp	109		Aa j	Ile	Gly	Ser	Ser 109		. Thr	Asn
Val	Ala 1100		Leu	ı Ser	Gln	Asr 110		le (	Gln	Gly	Ser	Asp 1110		Leu	Glu
Ala	Ile 1115		s Asp	Gly	Lys	Th: 112		lu I	Met	Ser	Leu	Asn 112		ı Asp	Gly
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Arg	Ile 1160		glu	ı Ile	· Val	116 116		is 1	Phe	Ala	Lys	Asp 1170		. Asn	Ser
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Ile	Gly 1205		ı Glu	ı Lys	Asp	Arg 121		al I	Lys	Ala	Tyr	Arg 121		Glu	Ile
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Asn	Ala 1235		: Asp	Thr	Gly	Asr 124		rg 1	Lys	Pro	Cys	Thr 124		ılle	Thr
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Thr	Ala 1265		ı Lev	ı Glu	ı Glu	. Val		λa ,	Val	Gly	Glu	Arg 127		. Leu	Pro
Asn	Ala 1280		туг	Ala	. Leu	. Asp		er '	Tyr	Ser	Val	Pro 1290		. Thr	Asp

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Ala	Pro 1415	Met	Ile	Asp	Ala	Leu 1420	Arg	Tyr	Met	Pro	Arg 1425	Ser	Asn	Gly
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Gln	Thr 1460	Gly	Trp	Met	Ile	Leu 1465	Gly	Phe	Glu	Glu	Pro 1470	Val	Gln	Ala
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Thr	Glu 1505	Ala	Pro	Ala	Pro	Ser 1510	Glu	Lys	Tyr	Thr	Ile 1515	Thr	Ala	Asn
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Thr	Ile 1580	Thr	Ala	Asn	Phe	Glu 1585	Arg	Ile	Glu	Val	Glu 1590	Asn	Glu	Gly
Trp	Val 1595	Gln	Thr	Glu	Asn	Gly 1600	Trp	Glu	Tyr	Tyr	Glu 1605	Asn	Gly	Gln
ГÀа	Val 1610	Val	Gly	Trp	Lys	Glu 1615	Val	Ser	Gly	Lys	Trp 1620	Tyr	Tyr	Phe
Glu	Glu 1625	Asn	Gly	Leu	Met	Gln 1630	Thr	Gly	Trp	Val	Phe 1635	Val	Asn	Asn
His	Trp 1640	Tyr	Tyr	Met	Asp	Gln 1645	Trp	Gly	Ala	Met	Cys 1650		Gly	Trp
Val	Ala 1655	Val	Asp	Gly	His	Trp 1660	Tyr	Tyr	Met	Asp	Gln 1665	Trp	Gly	Ala
Met	Cys 1670	Thr	Gly	Trp	Val	Ser 1675	Val	Asn	Gly	His	Trp 1680	Tyr	His	Met
Asp	Gln	Trp	Gly	Ala	Met	Gln	Thr	Gly	Trp	Ala	Leu	Val	Asp	Ser

												COII	CIII	ueu	
	1685	5				169	90				1	695			
Asn	Trp	_	ту1	. Le	ı Asr	170		sp G	Ly Se	er Me		la 710	Ile	Gly '	Trp
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< 400	)> SE	QUEI	ICE :	13											
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Ala	Val	Leu	Ala 20	Val	Thr	Ala	Ala	Gly 25	Val	Ser	Val	Pro	Ala 30	Leu	Ala
Gln	Gln	Ala 35	Val	Arg	Thr	Glu	Ser 40	Gln	Thr	Gln	Met	Ser 45	Ser	Asp	Pro
Glu	Leu 50	Val	Tyr	Val	Asn	Asn 55	Tyr	Ser	Ser	Thr	Ala 60	Gln	Arg	Ser	Gln
Asn 65	Phe	Asn	Ser	Asn	Trp 70	Lys	Phe	Tyr	Phe	Gly 75	Asp	Ala	Gly	Asn	Ala 80
Gln	Gly	Ala	Thr	Phe 85	Asp	Asp	Ser	Lys	Trp 90	Glu	Gln	Val	Ser	Leu 95	Pro
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His	Pro	Tyr	Gly	Tyr 165	Thr	Ser	Phe	Ser	Phe 170	Asp	Ile	Thr	Asp	Tyr 175	Val
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Pro	Ser	Ser 195	Arg	Trp	Tyr	Ser	Gly 200	Ser	Gly	Ile	Tyr	Arg 205	Asp	Val	Asp
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Thr 225	Thr	Pro	Asn	Leu	Glu 230	Thr	Glu	Lys	Gly	Ser 235	Thr	Val	Asn	Thr	Asp 240

Val	Thr	Ala	Thr	Val 245	Ala	Asn	Asp	Ser	Asp 250	Ala	Ala	ГÀа	Ser	Val 255	Ala
Val	Arg	His	Thr 260	Val	Phe	Pro	Lys	Asp 265	Gly	Ser	Ala	Asp	Gln 270	Ser	Ile
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Phe	Asn	Gln 435	Thr	Val	Ala	Ser	Asp 440	Asn	Glu	Ile	Leu	Gly 445	Ala	Thr	Asn
Gly	Glu 450	Thr	Trp	Ala	Gln	Phe 455	Asp	Leu	Glu	Ser	Met 460	Ile	Ser	Arg	Asp
Tyr 465	Asn	Ala	Pro	Ser	Val 470	Ile	Met	Trp	Ser	Leu 475	Gly	Asn	Glu	Val	Met 480
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Tyr	Asp	Asn 595	Ser	Asn	Val	Gly	Trp	Gly	Ala	Thr	Ala	Ser 605	Asn	Ala	Trp
Tyr	Thr 610	Val	Leu	Thr	Arg	Asp 615	Phe	Ala	Ala	Gly	Glu 620	Tyr	Val	Trp	Thr
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Ile	Asp	Thr	Ala	Gly	Phe	Ala	Lys	Asp	Ser	Tyr	Tyr	Phe	Tyr	Gln	Ser

-continued
Concinaca

_			660					665					670		
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Thr	Thr	Asp 755	Lys	Asn	Leu	Tyr	Leu 760	Thr	Trp	Asn	Val	Pro 765	Tyr	Ala	Asp
Gly	Thr 770	Val	Ser	Ala	Val	Ala 775	Tyr	Asn	Ser	Asn	Gly 780	Gln	Lys	Ile	Thr
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Leu	ГЛа	Ala	Ser	Ala 805	Asp	His	ГЛа	Lys	Ile 810	Ala	Ala	Asp	Gly	Glu 815	Ser
Leu	Ser	Tyr	Ile 820	Thr	Val	Asp	Val	Thr 825	Asp	Ala	Asn	Gly	Asn 830	Ile	Val
Pro	Asp	Ala 835	Glu	Asn	Arg	Val	Lys 840	Phe	Thr	Val	Glu	Gly 845	Asp	Gly	Glu
Leu	Val 850	Gly	Val	Asp	Asn	Gly 855	Ser	Ser	Pro	Asp	His 860	Asp	Ser	Tyr	Gln
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Ser	Thr	Lys	Glu	Ala 885	Gly	Thr	Ile	Thr	Val 890	Thr	Ala	Ser	Ala	Asp 895	Gly
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Val	Met 102		y Va∶	l Th:	r Phe	Pro 103		al Th	nr Ti	rp G		sp :	Lys i	Ala i	Ala
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The invention claimed is:

- 1. A method for producing a food product, the method comprising:
  - providing a substrate comprising lactose at a concentration of 5% w/w or greater;
  - adding a polypeptide to the substrate comprising lactose, the polypeptide having transgalactosylating activity, and the polypeptide selected from the group consisting of:
    - a. a polypeptide comprising an amino acid sequence having at least 90% sequence identity to the amino acid sequence of SEQ ID NO: 1,
    - a polypeptide comprising an amino acid sequence 50 having at least 90% sequence identity to the amino acid sequence of SEQ ID NO: 2,
    - c. a polypeptide encoded by a polynucleotide that hybridizes under high stringency conditions of 0.1 SSC (0.15 M NaCl, pH 7.0) at 65° C. with,
      - i) the nucleic acid sequence comprised in SEQ ID NO: 10 encoding SEQ ID NO: 1,
      - ii) the cDNA sequence of i), or
      - iii) the complementary strand of i) or ii),
    - d. a polypeptide encoded by a polynucleotide that hybridizes under high stringency conditions of 0.1 SSC (0.15 M NaCl, pH 7.0) at 65° C. with,
      - i) the nucleic acid sequence comprised in SEQ ID NO: 11 encoding SEQ ID NO: 2,
      - ii) the cDNA sequence of i), or
      - iii) the complementary strand of i) or ii),

- e. a polypeptide comprising a conservative substitution, deletion or insertion of in a sequence resulting in a sequence having at least 90% identity to SEQ ID NO:1, or
- f. a polypeptide comprising a conservative substitution, deletion or insertion of amino acids in a sequence resulting in a sequence having at least 90% identity to SEQ ID NO: 2,
- provided that the polypeptide of above items a, c, and e, at the most has a length of 1806 amino acids and provided that the polypeptide of above items b, d, and f at the most has a length of 1767 amino acids; and producing the food product that contains galacto-plices.
- producing the food product that contains galacto-oligossacharides.
- 2. A method of producing a dairy product, the method comprising:
  - providing a milk-based substrate comprising lactose at a concentration of 5% w/w or greater;
  - adding a polypeptide to the substrate comprising lactose, the polypeptide having transgalactosylating activity, and the polypeptide selected from the group consisting of:
    - a. a polypeptide comprising an amino acid sequence having at least 90% sequence identity to the amino acid sequence of SEQ ID NO: 1,
    - b. a polypeptide comprising an amino acid sequence having at least 90% sequence identity to the amino acid sequence of SEQ ID NO: 2,
    - c. a polypeptide encoded by a polynucleotide that hybridizes under high stringency conditions of 0.1 SSC (0.15 M NaCl, pH 7.0) at 65° C. with,

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- i) the nucleic acid sequence comprised in SEQ ID NO: 10 encoding SEQ ID NO: 1,
- ii) the cDNA sequence of i), or
- iii) the complementary strand of i) or ii),
- d. a polypeptide encoded by a polynucleotide that 5 hybridizes under high stringency conditions of 0.1 SSC (0.15 M NaCl, pH 7.0) at 65° C. with,
  - i) the nucleic acid sequence comprised in SEQ ID NO: 11 encoding SEQ ID NO: 2,
  - ii) the cDNA sequence of i), or
  - iii) the complementary strand of i) or ii),
- e. a polypeptide comprising a conservative substitution, deletion or insertion of in a sequence resulting in a sequence having at least 90% identity to SEQ ID NO:1, or
- f. a polypeptide comprising a conservative substitution, deletion or insertion of amino acids in a sequence resulting in a sequence having at least 90% identity to SEQ ID NO: 2,
- provided that the polypeptide of above items a, c, and e, 20 at the most has a length of 1806 amino acids and provided that the polypeptide of above items b, d, and f at the most has a length of 1767 amino acids; and producing the dairy product that contains galacto-oligossacharides.
- 3. The method of claim 1, further comprising treating the substrate with a hydrolysing beta-galactosidase.

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